Sediment Characterization Sampling and Analysis Plan (SAP) for the Levin-Richmond Terminal Corporation Berth A

Maintenance Dredging Program

Episode 1

Prepared for

Levin-Richmond Terminal Corporation
402 Wright Avenue
Richmond, CA 94804

Prepared by

Pacific EcoRisk
2250 Cordelia Road
Fairfield, CA 94534

November 2008

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ENVIRONMENTAL CONSULTING & TESTING

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Ms. Debra O'Leary
U.S. Army Corps of Engineers
San Francisco District
1455 Market Street
San Francisco, CA 94103-1398

November 5, 2008

Dear Ms. O'Leary:

On behalf of Mr. Jim Cannon of the Levin-Richmond Terminal Corporation (LRTC), please find enclosed three (3) copies of the Sampling and Analysis Plan "Sediment Characterization Sampling and Analysis Plan (SAP) for the Levin-Richmond Terminal Corporation Berth A" in support of the shipyard's maintenance dredging program. In addition, one copy of this SAP has been sent to the other DMMO participating agency representatives. This SAP has been prepared to support dredging of approximately 5,200 cubic yards of material from LRTC's Berth A.

LRTC, located in Point Richmond (CA) in the Richmond Inner Harbor Channel (Figures 1-1 and 1-2), is currently seeking 10-year permits/certification from the U.S. Army Corps of Engineers (USACE), the Bay Conservation and Development Commission (BCDC) and San Francisco Bay Regional Water Quality Control Board (RWQCB) for maintenance dredging of their Berth A. LRTC has contracted Pacific EcoRisk to assist in the preparation of a Sampling and Analysis Plan (SAP) in support of the first maintenance dredging episode under the new permits. LRTC is applying for the appropriate permits in parallel to this SAP.

Recent testing performed on LRTC Berth A sediments indicated that the sediments were suitable for disposal at the Montezuma Wetlands Project (Montezuma); it is anticipated that future dredged material will also be disposed at Montezuma as long as there is available capacity and sediment quality is of similar nature. However, since sediment quality may vary over time and in the event that other disposal options may be available based on the results of the chemical analysis of the sediments, this SAP covers testing for a variety of disposal site options so as to ensure flexibility for the LRTC maintenance-dredging program.

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If you have any questions, please give me a call at (707) 207-7761. I look forward to hearing from you.

Sincerely,

Jeffrey Cotsifas
President

cc (w/enc): Brian Ross, U.S. EPA
Brenda Goeden, BCDC
Beth Christian, SFRWQCB
George Isaac, CDFG
David Woodbury, NMFS
Donn Oetzel, SLC
Jim Cannon, LRTC

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List of Acronyms

ASTM	American Society for Testing and Materials
Bay	San Francisco Bay
BCDC	Bay Conservation and Development Commission
COC	Chain-of-custody
CV	Coefficient-of-variation
DGPS	Differential global positioning system
DMMO	Dredged Material Management Office
EMI	EnviroMatrix Inc.
ESC	Elutriate suitability concentrations
GPS	Global positioning system
IAA	Integrated alternatives analysis
ITM	Inland Testing Manual
LRTC	Levin-Richmond Terminal Corporation
mg/kg	Milligram per kilogram
MLLW	Mean lower low water
MRL	Method reporting limit
NOAA	National Oceanic and Atmospheric Administration
PAH	Polynuclear aromatic hydrocarbon
PCB	Polychlorinated biphenyls
PER	Pacific EcoRisk
QA/QC	Quality assurance/quality control
RPD	Relative percent difference
RWQCB	Regional Water Quality Control Board
SAP	Sampling and analysis plan
SF-DODS	San Francisco Deep Ocean Disposal Site
SLC	State Lands Commission
SOP	Standard operating procedures
SUAD	Suitable for undefined aquatic disposal
TEG	TEG Oceanographic Services
TOC	Total organic carbon
USACE	U.S. Army Corps of Engineers
USEPA	U.S. Environmental Protection Agency
µg/kg	Microgram per kilogram

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1. INTRODUCTION

The Levin-Richmond Terminal Corporation (LRTC), located in Richmond, CA, in the Richmond Inner Harbor Channel (Figures 1-1 through 1-3), is currently seeking 10-year permits/certification from the U.S. Army Corps of Engineers (USACE), the Bay Conservation and Development Commission (BCDC) and San Francisco Bay Regional Water Quality Control Board (SFRWQCB) for maintenance dredging of their Berth A. LRTC has contracted Pacific EcoRisk to assist in the preparation of a Sampling and Analysis Plan (SAP) in support of the first maintenance dredging episode under the new permits. LRTC is applying for the appropriate permits in parallel to this SAP. It is anticipated that maintenance dredging will be performed over multiple episodes over the course of the permit period.

To accommodate vessel transit and berthing, LRTC requires dredging of its Berth A area to a depth of –39.0 ft MLLW + 2.0 ft over-dredge; it is proposed that this area be sampled and tested to a total depth of –41.0 ft MLLW. The proposed maintenance depth and estimated volumes of dredged material for each area, including over-depth, are summarized in Table 1-1.

Table 1-1. Proposed maintenance dredging for the Levin-Richmond Terminal Corporation

Area	Permitted Depth (ft MLLW)	Estimated Volume (yds^3)	Over-depth (ft)	Estimated Volume (yds^3)	Maintenance Depth (ft MLLW)	Total Estimated Volume (yds^3)
Berth A	-39.0	1,780	+2	3,346	-41	5,126

Testing of LRTC Berth A sediments in 2005 indicated that the sediments were suitable for "deep-cell" disposal at the Montezuma Wetlands Project (Montezuma); it is anticipated that future dredged material will also be disposed of in "deep-cells" at Montezuma as long as there is available capacity and sediment quality is of similar nature. However, since sediment quality may vary over time and in the event that other disposal options, such as the Alcatraz Disposal Site (SF-11), the San Francisco Deep Ocean Disposal Site (SF-DODS), or Hamilton Wetlands, become viable options based on the results of the chemical analysis of the sediments, this SAP covers sampling and testing for a variety of disposal site options so as to ensure flexibility for the LRTC maintenance-dredging program.

The testing portion of the program will be performed in a Tiered process with the assessment of sediment chemical concentrations being performed prior to any other testing. If the analytical chemistry results indicate that sediments would be suitable for unconfined aquatic disposal or placement as cover material at a wetland re-use site, then the biological testing component of the

program, specific to the preferred alternative, will be implemented. Otherwise, "deep-cell" placement at the Montezuma Wetlands Project or other appropriate alternative will be pursued.

This Sampling and Analysis Plan (SAP) is being prepared in support of maintenance dredging in which the LRTC is proposing to dredge depositional material from Berth A, and has been developed in accordance with currently applicable guidance and establishes the general approach to sampling and assessment of sediments proposed for dredging. General sampling locations within Berth A are provided in Figure 1-4.

1.1 Objectives of the Sediment Investigation

The purpose of any sampling and testing program will be to evaluate the proposed dredged material to determine whether it will represent an adverse impact during removal operations and placement at a currently permitted disposal sites and/or future alternative disposal sites. The procedures for sediment sample collection, sample processing and preparation, physical and chemical analyses, biological testing and data analyses are presented in this SAP.

Guidance concerning necessary sampling and analytical protocols, quality assurance/quality control (QA/QC) procedures, and data interpretation used in preparation of this multi-year SAP is found in:

- Evaluation of Dredged Material Proposed for Ocean Disposal: Testing Manual (OTM; USEPA/USACE 1991);
- Evaluation of Dredged Material Proposed for Discharge in Waters of the U.S. – Testing Manual (ITM; USEPA/USACE 1998);
- Public Notice 01-1: Guidelines for Implementing the Inland Testing Manual in the San Francisco Bay Region;
- Public Notice 99-4: Proposed Guidance for Sampling and Analysis Plans (Quality Assurance Project Plans) for Dredging Projects within the USACE San Francisco District;
- Public Notice 93-2: Testing Guidelines for Dredged Material Disposal at San Francisco Bay Sites;
- The Dredged Material Management Office (DMMO) review process;
- Endangered Species Consultation for the Proposed Wetland Restoration Project at the Former Hamilton Airfield, City of Novato, Marin County, California. U.S. Fish and Wildlife Service Letter 1-1-0-F-0068 (USFWS 2005);
- San Francisco Bay Regional Water Quality Control Board Order No 00-061 Waste Discharge Requirements for: Levine-Fricke Restoration Corporation and Montezuma Wetlands LLC, Montezuma Wetlands Restoration Project, Solano County; and,
- Personal communication (e-mail) Rachel Bonnefil. Analytical Requirement for Montezuma August 24, 2007.

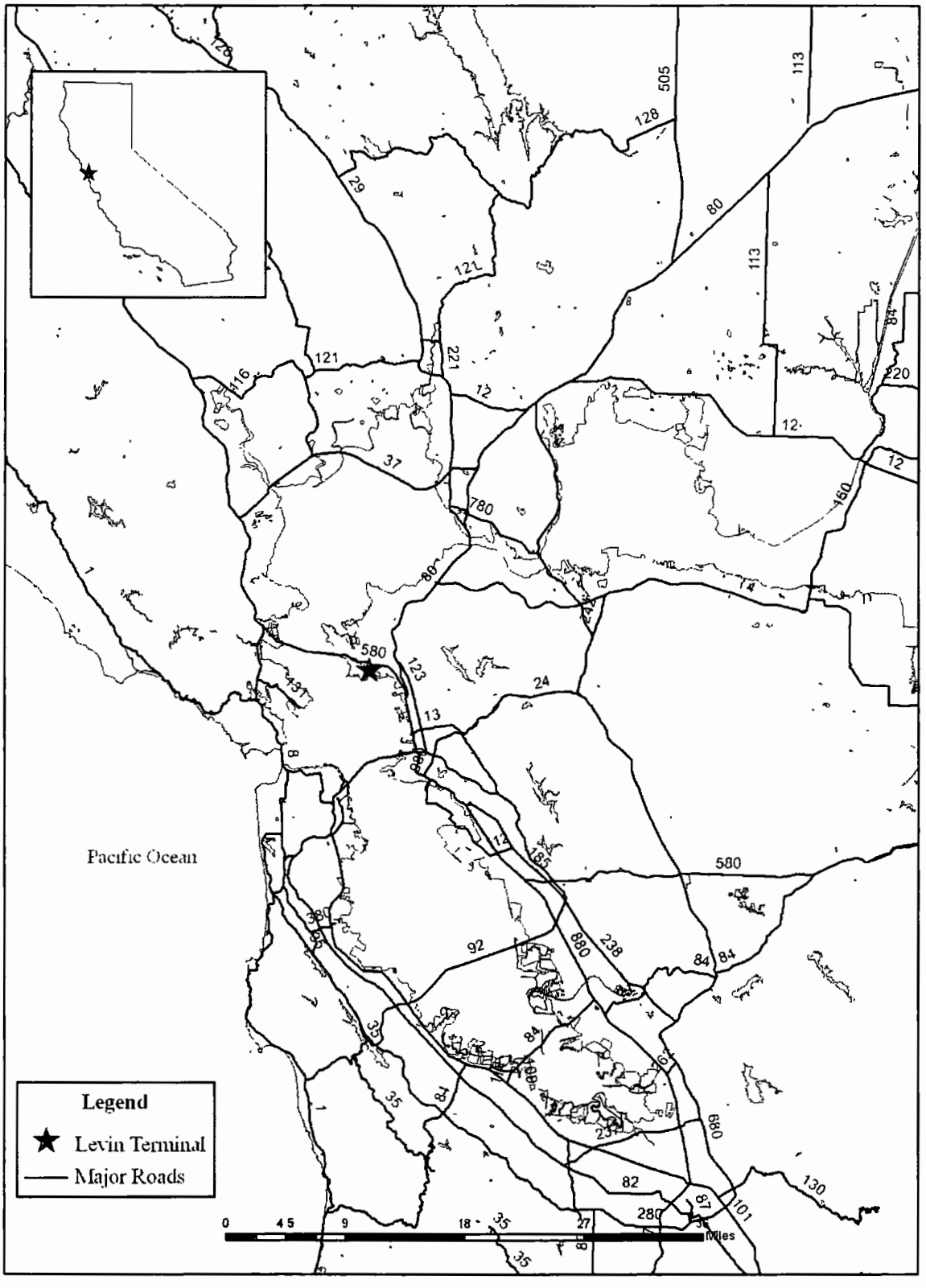


Figure 1-1. Location map: Levin-Richmond Terminal

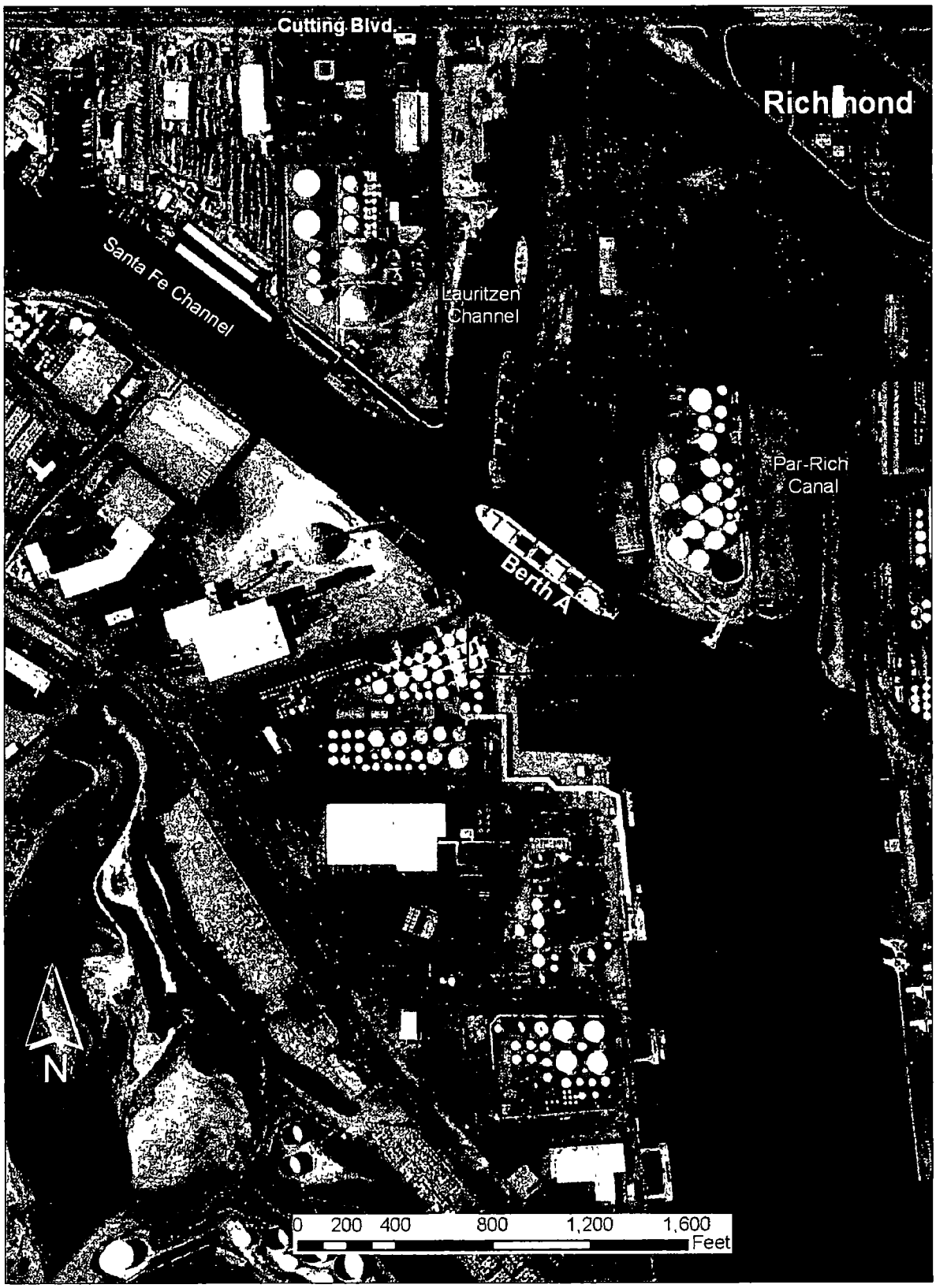


Figure 1-2. Vicinity Map #1: Levin-Richmond Terminal

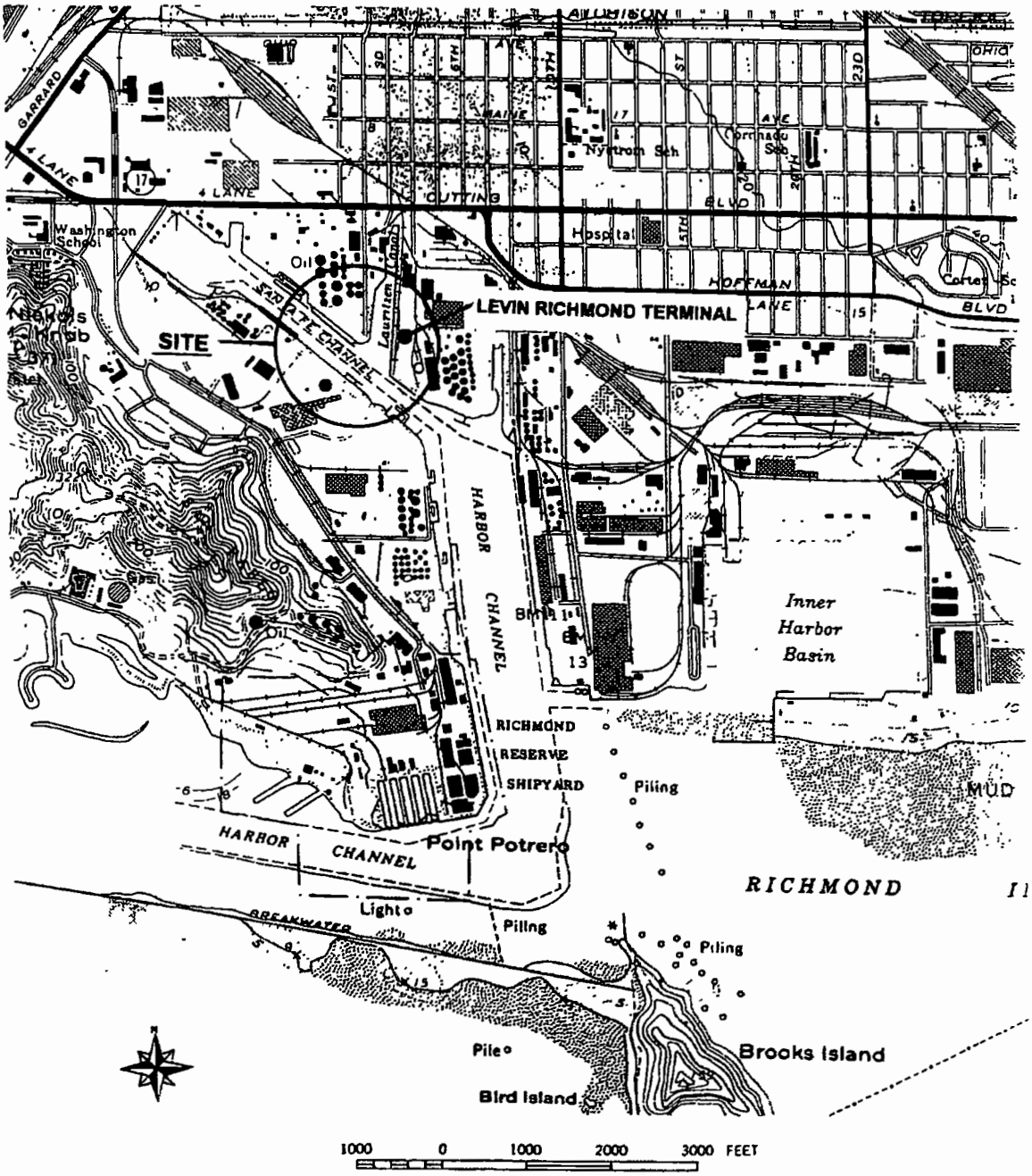
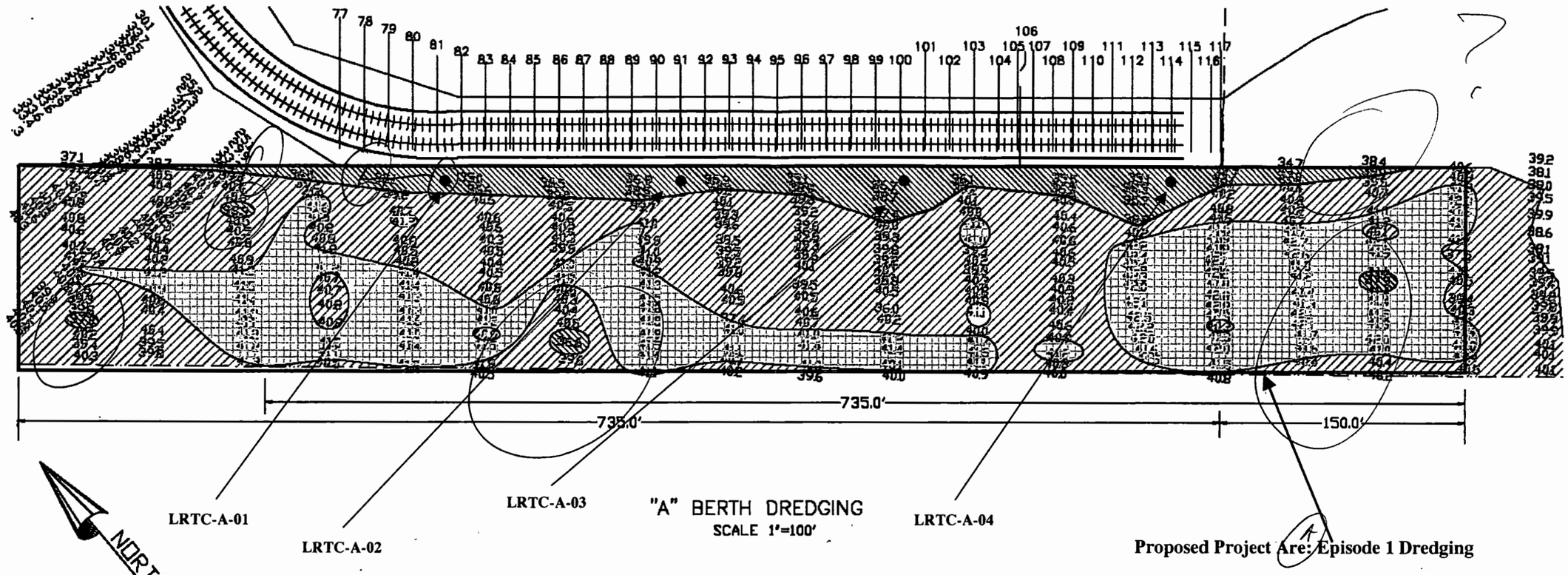


Figure 1-3. Vicinity Map #2: Levin-Richmond Terminal



LEGEND

<-39'	
-39' TO -41'	
>-41'	

DREDGING VOLUMES

TO -39' =1,780 yds

TO -41' =5,126 yds

NOTES:

1. BENCHMARK: NAIL AND SHINER SET IN WHARF. EL=11.63' MLLW.
2. BATHMETRIC DATA SHOWN REPRESENTS WATER DEPTHS AT A 0.0 TIDE; ALL ELEVATIONS SHOWN ARE NEGATIVE, UNLESS NOTED OTHERWISE.
3. SOUNDINGS TAKEN BY FATHOMETER ARE SHOWN TO THE NEAREST TENTHS OF A FOOT.
4. COORDINATE VALUES ARE BASED ON THE CALIFORNIA COORDINATE SYSTEM, ZONE III, NAD '83.
5. THE INFORMATION DEPICTED ON THIS MAP REPRESENTS RESULTS OF A HYDROGRAPHIC SURVEY MADE ON 08-29-08, AND INDICATES THE CONDITIONS AS OF THAT DATE.
6. SOUNDINGS WERE TAKEN USING AN ODOM HYDROTRAC ECHOSOUNDER; HORIZONTAL LOCATION BY LEICA RTK DGPS SOFTWARE UTILIZED IS COASTAL OCEANOGRAPHICS 'HYPACK'.
7. BACKGROUND LAND TOPOGRAPHIC DATA RECEIVED FROM LEVIN-RICHMOND TERMINAL.

Figure 1-4. Levin-Richmond Terminal Berth "A" Sample Locations

The results of all appropriate sediment analyses will be used to determine the suitability of the proposed sediments for "deep-cell" disposal at Montezuma; however; if the results of sediment chemical analysis indicates that the sediment may be suitable for either unconfined aquatic disposal or wetland reuse, then further testing will be pursued. Suitability for disposal at the SF-11 and SF-DODS disposal site will be determined by comparison to SF-11 and SF-DODS reference area databases. Suitability for placement at a wetland reuse site will be determined by compliance with site-specific requirements.

1.2 Overview of Field Activities and Lab Analyses

A total of 4 sediment cores will be collected from within LRTC Berth A using a vibra-corer (Figures 1-4). A sub-sample of the sediment from each core will be archived for subsequent analyses of the individual core sediment, if needed. Proportional aliquots of the sediment from the cores collected from each site will be composited; a sample of each composite sediment will be submitted for chemical and conventional analyses with biological testing being deferred until the results chemical analysis are determined. Based on these results, a determination of further testing to be performed, will be determined in consultation with the Dredged Material Management Office (DMMO). The results of these analyses will be used to determine the suitability of the proposed sediments for disposal at an approved disposal site.

1.3 DMMO Agency Review and Permitting

The federal and state agencies responsible for regulating dredged material programs in the San Francisco Bay area include:

- U.S. Environmental Protection Agency (USEPA) Region 9,
- U.S. Army Corps of Engineers (USACE),
- San Francisco Regional Water Quality Control Board (SFRWQCB),
- San Francisco Bay Conservation and Development Commission (BCDC), and
- State Lands Commission (SLC).

Representatives from these agencies comprise the DMMO.

LRTC is applying for a new permit or/certification from each of the DMMO Agencies to conduct maintenance dredging in at their Berth A. Collectively, Episode 1 for this project entails dredging of approximately 5,200 cubic yards of material; it is anticipated that up to 200,000 cubic yards will be removed over a 10-year period.

2. PROJECT MANAGEMENT AND RESPONSIBILITIES

2.1 Program and Field Activities

Mr. Jim Cannon (of LRTC) will be the Project Manager. The Sampling and Analysis Project Manager will be Mr. Jeff Cotsifas (of Pacific EcoRisk [PER]), assisted by Dr. Scott Ogle. Mr. Cotsifas will be responsible for overall project coordination, including production of all project deliverables, collection and submittal of environmental samples to the designated laboratories for chemical and physical analyses, and administrative coordination to assure timely and successful completion of the project. Mr. Cotsifas will also be responsible for all decisions concerning sample collection, for QA/QC oversight, and ensuring that appropriate protocols for decontamination, sample preservation, and holding times are observed. Mr. Cotsifas will be involved in all aspects of this project, including preparation, and approval of the SAP, and review and interpretation of all analytical results; Dr. Ogle will be involved in review and interpretation of all analytical results. The project management organization is illustrated in Figure 2-1.

All field activities will be performed under the direction of Mr. Cotsifas. Sediment cores will be collected by TEG Oceanographic Services (TEG) and PER. During collection of cores, the sampling vessel will be staffed with a captain, operating crew, and 2 field scientists. Mr. Mark Mertz of TEG will captain the sampling vessel, and will be responsible for location control and positioning, and providing all coring devices and operating crew. PER will supply a Field Manager and Field Scientist.

2.2 Project Management

A Laboratory Project Manager will be appointed from each laboratory. Laboratory Project Managers will provide analytical support and will be responsible for ensuring that all laboratory analyses meet the project data quality objectives and other specifications required by the ITM, regional guidance, and the DMMO review process. The Laboratory Project Managers are as follows:

Project Management and Bioassay Testing:
Mr. Jeffrey Cotsifas
Pacific EcoRisk
2250 Cordelia Road
Fairfield, CA 94534
Telephone: (707) 207-7760
Facsimile: (707) 207-7916

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San Diego, CA 92123
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Vibracore Sampling Vessel Operation:
TEG Oceanographic Services
216 Florence Drive
Santa Cruz, CA 95061
Telephone: (831) 684-2749
Facsimile: (831) 684-2748

The contract laboratories and vessel operators are expected to meet the following minimum technical requirements as specified in their negotiated subcontracts with PER:

1. Adherence to the methods outlined in the SAP, including those methods referenced for each analytical procedure, as per ITM, PN-01-01, and DMMO requirements;
2. Deliver electronic data files as specified;
3. Meet all reporting requirements;
4. Implement and comply with QA/QC procedures required by ITM and DMMO guidelines;
5. Allow PER to perform laboratory and data audits; and,
6. Meet turnaround times for deliverables.

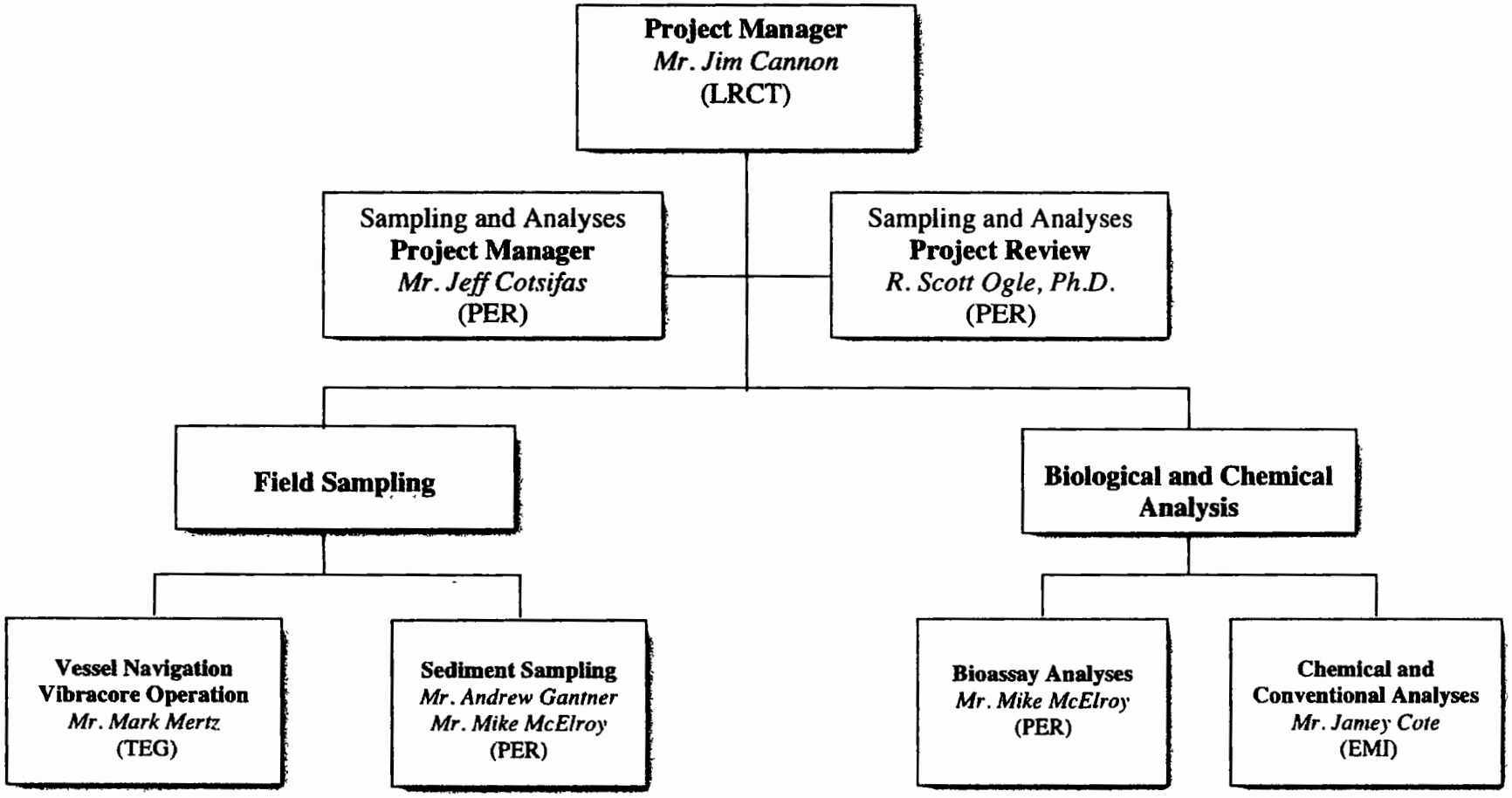

Figure 2-1. Project Organizational Chart

3. REVIEW OF EXISTING DATA

3.1 Site History

The LRTC Loading Terminal is located in the Richmond Inner Harbor in Richmond, CA. The eastern end of the facility is bordered by 8th Street. Manson Construction is located northwest of the terminal facility; Kinder Morgan and ConocoPhillips are located directly across the channel.

A portion of the LRTC property is located on what is currently known as the United Heckathorn Superfund site. The site is in an industrial area dominated by petroleum and shipping terminals. From 1947 to 1966, several operators, including the R.J. Prentiss Company, Heckathorn and Company, United Heckathorn, United Chemetrics, and Chemwest Incorporated (collectively referred to as "United Heckathorn") used the site to formulate and package pesticides. No chemicals were manufactured on site. United Heckathorn received technical grade pesticides from chemical manufacturers, ground them in air mills, and mixed them with other ingredients such as clays or solvents, after which they were packaged for final use in liquid or powder formulations. Although many pesticides were handled by United Heckathorn, DDT accounted for approximately 95% of its operations. United Heckathorn went bankrupt and vacated the site in 1966. Between 1966 and 1970 the United Heckathorn buildings were demolished and cleared from the site. In the 1970s, the site was used primarily for bulk storage. In 1981, the Levin-Richmond Terminal Corporation purchased the property and currently operates a bulk shipping facility at the site (USEPA 2002a).

3.1.1 Storm Drain, Spills and Discharges

To LRTC's knowledge, there have been no spills or other environmental events on their property that would materially change the quality of the Terminal's Berth A sediments. All storm drains enter into a canal north of Site 2. The LRTC storm drains are regulated by the RWQCB under a NPDES permit; all discharges from these drains have met NPDES permit requirements.

3.2 Recent Testing History

Under a previous permit (USACE 29762S) or certification from each of the DMMO Agencies, maintenance dredging has was performed at LRTC Berth A and the adjacent Pacific Atlantic (formally Shore) Terminal berth; due to elevated total DDT levels, this material was placed in deep-cells at the Montezuma Wetlands Site. The results of this testing performed in 2005 (PER 2006a, 2006b, 2006c, 2006d) are presented below in Sections 3.2.1 and 3.2.2. Testing results for previous maintenance dredging events performed at this facility prior to 2005 were not available; however, there is reference in a previous permit (USACE 24314S) that approximately 7500 cubic yards of dredged material removed form Berth A in 2000 was placed at the Port of Richmond Shipyard #3 and used as sub-base for a new parking lot.

3.2.1 Recent Testing for Levin-Richmond Terminal Berth A- October 2005

This sediment was ~44% total solids and contained 1.72% TOC, which is typical for San Francisco Bay. Grain size analyses indicated that the sediment was 76.2% silts and clays, 18.2% sand, and 3.3% gravel.

All metal analytes were generally similar to ambient bay concentrations (SFRWQCB, 1998). The total organotin concentration was 45.7 μg/kg. The total PAH concentration was 4,664 μg/kg. Total DDT concentrations ranged from 274-462 μg/kg with dieldrin and heptachlor epoxide measured at 8.7 and 1.7 μg/kg, respectively; all other organochlorine pesticides and PCB Aroclors were below their respective detection limits.

Biological testing indicated that there was no toxicity to amphipods or polychaetes; all elutriate samples were below the limited permissible concentration (LPC) for sediment disposal at in-bay sites.

Based on the above testing results, the DMMO determined that all of the sediments were suitable for disposal at the Montezuma Wetlands Projects deep cells; summary tables of the analytical chemistry and bioassay results for each testing event are presented in Appendix B.

3.2.2 Recent Testing for Pacific Atlantic (formerly Shore) Terminal - October 2005

This sediment was ~45% total solids and contained 1.14% TOC, which is typical for San Francisco Bay. Grain size analyses indicated that the sediment was 89.1% silts and clays, 13.3% sand, and 0% gravel.

All metal analytes were generally similar to ambient bay concentrations (SFRWQCB, 1998). The total organotin concentration was 29 μg/kg. The total PAH concentration was 110.4 μg/kg. Total DDT concentrations ranged from 140-290 μg/kg with dieldrin, endosulfan II, endrin ketone, and heptachlor epoxide measured at 3.4, 3.1, 1.4 and 1.2 μg/kg, respectively; all other organochlorine pesticides and PCB Aroclors were below their respective detection limits.

Biological testing indicated that there was no toxicity to amphipods or polychaetes; all elutriate samples were below the limited permissible concentration (LPC) for sediment disposal at in-bay sites.

Based on the above testing results, the DMMO determined that all of the sediments were suitable for disposal at the Montezuma Wetlands Projects deep cells; summary tables of the analytical chemistry and bioassay results for each testing event are presented in Appendix B.

4. SAMPLING PROGRAM: SEDIMENT COLLECTION AND HANDLING

4.1 Sampling Platform

TEG will provide the sampling vessel and all equipment necessary for the safe operation of the boat to support sampling operations. The sampling vessel is 35-ft long trawler vessel with a 4-ton belt hydraulic crane for deploying and retrieving sampling equipment; operation of the sampling vessel will be the responsibility of Mr. Mark Mertz. The vessel is powered by twin V12 diesel engines, has an AC/DC electrical system and approximately 35 x 20 ft^2 of clear aft deck work space for processing samples. The vessel conforms to U.S. Coast Guard safety standards.

Collection of sediment cores will be performed by both TEG and PER Field Scientists. Sediment cores will be collected and stored in appropriate sample containers on board the vessel.

4.2 Navigation and Vertical Control

Location control will be the responsibility of the boat captain(s) and will be accomplished using a differential global positioning system (DGPS). The navigation systems will be calibrated to a known survey monument in the project area. The navigation system will be used to guide the vessel to predetermined core sample locations and to identify the exact sampling location where the corer strikes the bottom. The required accuracy for horizontal positioning is ± 3 m.

Upon locating the sampling position, station depth will be measured using an on-board calibrated fathometer or a lead line, and tidal elevation will determined relative to harbor datum MLLW. The tidal elevation will be subtracted from the measured depth to determine the sediment surface elevation relative to MLLW. All vertical elevations will be report to the nearest foot relative to zero (0) ft MLLW, harbor datum.

In the event that the DGPS is not functioning properly because of local interference, station locations will be positioned using a laser range finder to record the perpendicular distance from at least two stationary markers located within the harbor. Interference with DGPS is not expected to be a problem at this location.

4.3 Station Locations

The objective of the sampling station selection and the subsequent compositing design is to provide samples that represent, as accurately as possible, the physical, chemical, and toxicological characteristics of the sediments to be dredged. Results of the most recent bathymetric survey were used to assist in choosing core sample stations (Figures 1-5). Sampling locations were chosen in areas that were representative in depth of the surface sediment above the proposed dredging depth at intervals within the proposed dredge limits to provide appropriate general coverage (Tables 4-1).

Table 4-1. Dredge Episode 1: Locations of sampling stations and estimated core depths

Sample ID	Latitude* (deg-min-sec)	Longitude* (deg-min-sec)	Mudline Elevation (ft MLLW)	Proposed Project Depth + Over-Depth (ft MLLW)	Estimated Core Length (ft)
LRT-S01-01	37° 55' 10.10"	122° 22' 00.78"	-33.0	-41	8.0
LRT-S01-02	37° 55' 08.90"	122° 21' 59.07"	-36.5	-41	4.5
LRT-S01-03	37° 55' 08.21"	122° 21' 57.74"	-36.7	-41	4.3
LRT-S01-04	37° 55' 07.68"	122° 21' 56.83"	-33.1	-41	7.9

*State Plane Coordinate System, California Zone 3, NAD 83

4.4 Collection of Sediment Core Samples

The sediment core sampling procedure is summarized in this section. Greater detail is provided in the Standard Operating Procedure (SOP) for sediment core collection (Appendix C).

All samples will be collected using an appropriate coring device. All cores will be collected to the dredge depth, or refusal. Upon completion of core penetration at a station, the position will be recorded and the sampler recovered.

Once the corer is on deck, the sediment core will be extracted from the corer barrel. The core will be examined to determine compliance with acceptability criteria as follows:

1. The core penetrated and retained material to project depth, or to refusal,
2. Cored material does not extend out the top of the core tube or contact any part of the sampling apparatus at the top of the core tube,
3. There are no obstructions in the cored material that might have blocked the subsequent entry of sediment into the core tube, resulting in incomplete core collection.

If core acceptance criteria are not achieved, the core will be rejected and the procedure repeated until acceptance criteria are met. If 3 repeated attempts within 25-50 ft in either direction of the proposed location do not yield a core that meets the appropriate acceptance criteria, the Sampling and Analysis Project Manager or field lead will select an alternate station of similar representability.

4.4.1 Collection of Site Water

Ambient surface water will be collected from within the DU areas for use in preparing the sediment elutriate for biological testing, should biological testing prove necessary. Briefly, site water will be collected from approximately 3 ft below the surface using a battery-operated peristaltic pump fitted with Tygon tubing. Site water will be "pre-pumped" through the tubing for approximately 3 minutes before the sample is collected. Water will then be pumped into a 10-

L polypropylene carboy, with the carboy being pre-rinsed 3 times with site water before the site water sample is collected. After the site water samples are collected, the carboys will be sealed, labeled, and stored on ice, until delivered to the bioassay laboratory.

4.4.2 Collection of Reference Sediments

Disposal site reference sediments will be not be collected as part of this testing program; existing reference site databases for the SF-11 and SF-DODs disposal sites will be utilized in the event an evaluation for unconfined aquatic disposal is performed.

4.5 On-Board Sample Processing and Labeling

Individual cores will be extruded and placed into food-grade polyethylene bags on board the sampling vessel. Physical characteristics of each core will be noted on the individual sediment core collection log. Aboard the vessel, samples will be temporarily stored on ice (or frozen "blue ice") within insulated coolers.

4.5.1 Station and Sample Identification

Each individual sediment core and composite sediment sample will be assigned a unique alphanumeric identifier using the format described below:

- The first 3 characters will identify the area e.g., LRT = Levin-Richmond Terminal,
- The next character will identify the Site, e.g., A = Berth A,
- The next two characters will be used to identify:
  1) the coring location, and
  2) the sequence of collection from that particular site.

  For coring locations and respective individual samples, these two characters will be 01, 02, 03, and 04.

Using this protocol, the individual station core samples for Site 1 will be identified as LRT-A-01, LRT-A-02, LRT-A-03, and LRT-A-04.

4.6 Field Equipment Decontamination Procedure

The deck of the vessel will be rinsed clean with site water between stations. All sampling equipment coming in contact with collected sediments will be decontaminated between stations using the following procedures:

1. Rinse with site water and wash with scrub brush until free of sediment,
2. Wash with phosphate-free biodegradable soap solution,
3. Rinse with site water taken from 3 ft below the surface.

Any sampling equipment that cannot be properly cleaned will not be used for subsequent sampling activity.

Acid- or solvent-washing will not be used in the field due to safety considerations and problems associated with rinsate disposal. Residue of acids and solvents on sampling equipment may affect sample integrity for chemical testing. The use of acids or organic solvents on the deck of a vessel may pose a safety hazard to the crew.

4.6.1 Waste Disposal

All sediment remaining on deck after sampling will be washed overboard at the collection site prior to moving to the next sampling station. All disposable sampling materials and personnel protective equipment used in sample processing, such as disposable coveralls, gloves, and paper towels, will be placed in heavy-duty garbage bags or other appropriate containers. Disposable supplies will be removed from the vessel by sampling personnel and placed in a normal refuse container for disposal as solid waste.

4.7 Field Data Recording

The Sampling and Analysis Project Manager, or his designee, will maintain a field logbook. The field logbook will provide a description of all sampling activities (including documentation of all samples collected for analysis), conferences associated with field sampling activities, sampling personnel, weather conditions, and a record of all modifications to the procedures and plans identified in this SAP. The field logbook is intended to provide sufficient data and observations to enable readers to reconstruct events that occurred during the sampling period.

Core collection log sheets will be completed for each sediment core. In addition to standard entries of personnel, date, and time, the log sheet will also include information regarding station coordinates, core penetration, and physical characteristics of the sediment such as texture, color, odor, stratification, and sheens.

4.8 Laboratory Sample Processing/Compositing Plan

Compositing of individual cores will be performed at the PER laboratory facility in Fairfield, CA. The sediment from each individual core will be individually homogenized in a stainless-steel bowl or high-density polyethylene (HDPE) container, whichever can accommodate the collected volume. A 500-mL sub-sample of each individual core will be archived to allow for additional chemical analyses, if necessary (archived samples will be stored frozen at –20 ± 10°C for up to one (1) year after sample collection). Representative portions of the remaining homogenized sediment from each of the cores will be proportionally combined to form a homogenized Berth A site composite sample.

For the samples being shipped to the analytical laboratories, sample labels will be filled out with an indelible-ink pen and affixed to the sample containers. Each label will contain the project number, sample identification number, preservation technique, requested analyses, date and time of collection and preparation, and initials of the person preparing the sample. To protect the

information on the sample labels, clear tape will be placed around the labeled sample containers. Appropriate aliquots of the homogenized Berth A composite samples will be placed into the sample containers, which will then be placed into a sample freezer and frozen until shipped, with the exception of sediment samples slated for grain size analysis, which will be stored at 4°C.

A 500-mL aliquot of the homogenized site composite will be archived as described above. The remaining homogenized site composite sediment will be stored at 4°C for potential subsequent biological testing, if required. The remaining sediments from each of the individual cores will also be stored at 4°C.

4.9 Sample Shipping

Prior to shipping to the analytical laboratory, sample containers will be wrapped in bubble wrap and securely packed inside a cooler with ice packs or crushed ice. A temperature blank will be included in each cooler. The original signed COC forms will be placed in a sealed plastic bag and taped to the inside lid of the cooler. Appropriate packaging tape will be wrapped completely around the cooler. A '*This Side Up*' arrow label will be attached on each side of the cooler, a '*Glass-Handle with Care*' label will be attached to the top of the cooler, and the cooler will be sealed with custody seals on both the front and the back lid seams.

Sediment samples will be shipped by overnight delivery. Each Laboratory Project Manager at each laboratory will ensure that appropriate chain-of-custody (COC) protocol is followed. The respective laboratory QA Officers will measure and record the temperature of the temperature blank included in each cooler and will specifically note any coolers that do not contain ice packs or are not sufficiently cold upon receipt.

The sub-contracting analytical laboratories will not dispose of any samples for this project until notified by PER in writing.

4.9.1 Chain-of-Custody (COC) Protocol

COC procedures will be followed for all samples throughout the collection, handling, and analyses activities. The Sampling and Analysis Project Manager, or a designee, will be responsible for all sample tracking and COC procedures. This person will be responsible for final sample inventory, maintenance of sample custody documentation, and completion of COC forms prior to transferring samples to the analytical laboratory. A COC form will accompany each cooler of samples to the respective analytical laboratories. Each person who has custody of the samples will sign the COC form; a copy of the COC form will be retained in the project file.

Each Laboratory Project Manager will ensure that COC forms are properly signed upon receipt of the samples and will note questions or observations concerning sample integrity on the COC forms. The Laboratory Project Manager will contact the Sampling and Analysis Project

Manager, or designee, immediately if discrepancies between the COC forms and the sample shipment are discovered.

5. LABORATORY ANALYSES

The testing portion of the program will be performed in a Tiered process with the assessment of sediment chemical concentrations being performed prior to any other testing. If the analytical chemistry results indicate that sediments would be suitable for unconfined aquatic disposal or placement as cover material at a wetland re-use site, then the biological testing component of the program, specific to the preferred alternative, will be implemented. Otherwise, "deep-cell" placement at the Montezuma Wetlands Project or other appropriate alternative will be pursued.

Chemical and conventional analyses and biological testing will be performed on composite samples to determine the suitability of the proposed dredged materials for unconfined aquatic disposal at the SF-11 or SF-DODS disposal sites, or a wetland reuse site (i.e., Hamilton Wetlands, Montezuma Wetlands Restoration Site) in the event that one is available and is determined to be the appropriate alternative.

5.1 Chemical and Conventional Analyses

All sediment chemical and conventional analyses will be conducted in accordance with ITM, OTM and DMMO guidelines. A brief summary of the proposed analyses of bulk sediment for different disposal options is presented below in Sections 5.1.1-5.1.3. A detailed list of each analysis, the analytical methods to be used, and the targeted reporting limits for the evaluation of sediments for SF-9 and SFDODS, Hamilton Wetlands, and Montezuma Wetlands are presented below in Tables 5-1 and 5-2, respectively. All samples will be maintained according to the appropriate holding times and temperatures for each analysis (presented in Appendix D).

5.1.1 Chemical Analyses of Sediments – In-Bay/Ocean Disposal Requirements

Chemical analyses will be performed on each of the sediments. All sediment analytical results will be presented on a dry weight basis (e.g., mg/kg or μg/kg, dry wt). Matrix spikes and sample duplicate analyses will be performed on the site sample. The chemical analyses to be performed are presented below in Table 5-1.

5.1.2 Chemical Analyses of Sediments – Montezuma Wetlands Requirements

The chemical analyses described in Sections 5.1.1 and 5.1.2 will cover all required analyses for potential disposal at the Montezuma Wetlands Restoration site for use as both cover and non-cover material. If the results of sediment analysis identify a contaminant(s) at elevated levels, a Wet Extraction Test (WET) using deionized water (DI) will be performed and analysis for the chemical of concern in the extract will be performed. All WET-DI analytical results will be presented in μg/L or mg/L. The chemical analyses to be performed will be dependent on the identification of a contaminant of concern at levels not suitable for unconfined aquatic disposal.

Table 5-1. In-bay/ocean disposal: List of analytes, methods, and targeted reporting limits.

Analyte	Method Reference	Targeted Reporting Limit (dry weight basis)
Metals		
Arsenic	EPA 6020	2 mg/kg
Cadmium	EPA 6020	0.3 mg/kg
Chromium	EPA 6020	5 mg/kg
Copper	EPA 6020	5 mg/kg
Lead	EPA 6020	5 mg/kg
Mercury	EPA 7471A	0.02 mg/kg
Nickel	EPA 6020	5 mg/kg
Selenium	EPA 7740	0.1 mg/kg
Silver	EPA 6020	0.2 mg/kg
Zinc	EPA 6020	1 mg/kg
Butyltins		
Mono-Butyltin	Krone 1989	10 µg/kg
Di-butyltin	Krone 1989	10 µg/kg
Tri-butyltin	Krone 1989	10 µg/kg
Tetra-butyltin	Krone 1989	10 µg/kg
Pesticides		
Aldrin	EPA 8081B	2 µg/kg
a-*BHC*	EPA 8081B	2 µg/kg
b-BHC	EPA 8081B	2 µg/kg
g-BHC (Lindane)	EPA 8081B	2 µg/kg
d-BHC	EPA 8081B	2 µg/kg
Chlordane	EPA 8081B	20 µg/kg
2,4'-DDD	EPA 8081B	2 µg/kg
2,4'-DDE	EPA 8081B	2 µg/kg
2,4'-DDT	EPA 8081B	2 µg/kg
4,4'-DDD	EPA 8081B	2 µg/kg
4,4'-DDE	EPA 8081B	2 µg/kg
4,4'-DDT	EPA 8081B	2 µg/kg
Total DDT	EPA 8081B	2 µg/kg
Dieldrin	EPA 8081B	2 µg/kg
Endosulfan I	EPA 8081B	2 µg/kg
Endosulfan II	EPA 8081B	2 µg/kg
Endosulfan sulfate	EPA 8081B	2 µg/kg
Endrin	EPA 8081B	2 µg/kg
Endrin aldehyde	EPA 8081B	2 µg/kg
Heptachlor	EPA 8081B	0.3 µg/kg
Heptachlor epoxide	EPA 8081B	0.3 µg/kg
Toxaphene	EPA 8081B	20 µg/kg

Table 5-1 (cont.) In-bay/ocean disposal: List of analytes, methods, and targeted reporting limits.

Analyte	Method Reference	Targeted Reporting Limit (dry weight basis)
PCBs		
Aroclor 1016	EPA 8082	20 µg/kg
Aroclor 1221	EPA 8082	20 µg/kg
Aroclor 1232	EPA 8082	20 µg/kg
Aroclor 1242	EPA 8082	20 µg/kg
Aroclor 1248	EPA 8082	20 µg/kg
Aroclor 1254	EPA 8082	20 µg/kg
Aroclor 1260	EPA 8082	20 µg/kg
PAHs		
Acenaphthene	EPA 8270C	20 µg/kg
Acenaphthylene	EPA 8270C	20 µg/kg
Anthracene	EPA 8270C	20 µg/kg
Benz(a)anthracene	EPA 8270C	20 µg/kg
Benzo(a)pyrene	EPA 8270C	20 µg/kg
Benzo(b)fluoranthene	EPA 8270C	20 µg/kg
Benzo(g,h,i)perylene	EPA 8270C	20 µg/kg
Benzo(k)fluoranthene	EPA 8270C	20 µg/kg
Chrysene	EPA 8270C	20 µg/kg
Dibenz(a,h)anthracene	EPA 8270C	20 µg/kg
Fluoranthene	EPA 8270C	20 µg/kg
Fluorene	EPA 8270C	20 µg/kg
Indeno(1,2,3-cd)pyrene	EPA 8270C	20 µg/kg
Naphthalene	EPA 8270C	20 µg/kg
Phenanthrene	EPA 8270C	20 µg/kg
Pyrene	EPA 8270C	20 µg/kg
Grain Size	ASTM 1992	0.1
Total Solids	Method 160.3	0.10%
Total Organic Carbon (TOC)	Method 415.1	0.10%

NOTES: µg/kg – microgram/kilogram
mg/kg – milligram/kilogram
PAH - polycyclic aromatic hydrocarbon
PCB - polychlorinated biphenyl

5.1.3 Chemical Analyses of Sediments – Hamilton Wetlands Requirements

In addition to the chemical analyses described in Sections 5.1.1, the following analyses (Table 5-2) will need to be performed to evaluate the suitability of sediments for potential disposal at the Hamilton Wetlands Restoration site for use as cover material. All sediment analytical results will be presented on a dry weight basis (e.g., mg/kg or µg/kg, dry wt).

Table 5-2. Hamilton Wetlands requirements: Additional list of analytes, methods, and targeted reporting limits.

Analyte	Reference Method	Targeted Reporting Limit (dry weight basis)
Metals		
Barium	EPA 6020	190 mg/kg
Beryllium	EPA 6020	1.03 mg/kg
Boron	EPA 6020	36.9 mg/kg
Cobalt	EPA 6020	27.6 mg/kg
Manganese	EPA 6020	943 mg/kg
Vanadium	EPA 6020	118 mg/kg
Organics		
Pentachlorophenol	EPA 8270 or EPA 8041	17 μg/kg
Phenol	EPA 8270	130 μg/kg
TPH-diesel/motor oil	EPA 8015d	144,000 μg/kg
TPH-gasoline/JP-4	EPA 8015d	12,000 μg/kg
BHC, total	EPA 8081B	0.99 μg/kg
Methoxychlor	EPA 8081B	90 μg/kg
Dioxins (total TCDD TEQ)	EPA 8290	0.02 μg/kg

5.2 Biological Testing

Toxicity and bioaccumulation tests are conducted (according to DMMO regional guidance and appropriate test protocol (i.e., ASTM Methods)) to determine whether anthropogenic contaminants of concern are present at concentrations that are toxic to biota, and whether removal of the sediment from the site and subsequent disposal at an unconfined aquatic disposal site or as wetland cover at an upland site poses a risk of toxicity to resident organisms. Benthic (whole sediment) and water column (sediment elutriate) toxicity tests, and sediment bioaccumulation tests will be conducted for each composite sediment. In addition, benthic toxicity tests will be performed on the test organisms' "Home" sediments and/or a Control sediment.

Test species selection and test procedures are discussed in the following sections. If the species proposed for testing are not available, or if the DMMO requests testing with different species, an appropriate alternative species will be selected from ITM/OTM Tables 11-1, 11-2, or 12-1. Summaries of test conditions for biological testing are presented in Appendix D.

5.2.1 Benthic Sediment Toxicity Testing

Benthic tests are conducted to evaluate the potential adverse toxicological impacts of dredged materials on the benthic community. These tests involve exposing organisms to test sediments and comparing the test organism responses with those exposed to the Control/reference sediments/reference site database. The 2 species proposed for benthic testing (the amphipod, *Ampelisca abdita,* and the polychaete, *Neanthes arenacoedentata*) exhibit 3 functional characteristics that represent important ecological usages of the benthic habitat: filter feeding, deposit feeding, and burrowing.

These tests will be performed using ASTM methods E1367-99 and E1611-00 for the amphipods and polychaetes, respectively. Ammonia and sulfide concentrations will be monitored in sediments immediately prior to setting up of each round of tests. If the ammonia or total sulfide concentrations in the bulk sediment interstitial waters (porewaters) exceed the recommended concentrations of 15 mg/L total ammonia (PN 01-01), or the calculated target value for the total sulfide (<0.56 mg/L at pH 7.5 [Knezovich et al., 1996]), then pre-test water exchanges (purging) will be required in order to reduce the ammonia and/or sulfide concentrations. In addition, if sediment porewater salinity is <25 ppt, salinity adjustment will be performed to bring the porewater salinity to >25 ppt.

If purging is necessary, it will begin immediately and will be applied to all replicates for all treatments including the negative control and reference sediments. Ammonia or sulfides will be purged by manually exchanging the overlying seawater in each test chamber twice daily. Once all total ammonia concentrations are at or below 15 mg/L, and/or total sulfide concentrations are below the calculated target value, the sediment test replicates will be loaded with test organisms and the tests will be initiated. Overlying water ammonia and/or sulfide concentrations will be monitored at test initiation (Day 0) and termination (Day 10). Salinity, pH, and temperature of the overlying water will also be measured at the test initiation and termination so that the un-ionized ammonia concentration can be calculated.

5.2.1.1 Amphipod Solid-Phase Survival Bioassay – One of the benthic test species will be the tube-dwelling amphipod *Ampelisca abdita*, with test organisms being collected from Narragansett, RI, or from San Francisco Bay, depending upon availability. All of the amphipods used in the project will be from one location to control for potential geographical genetic variability. Native "Home" control sediment will also be obtained from the amphipod collection site.

Amphipod tests will be conducted as 10-day (acute) static exposures, with 5 replicates per treatment. Each replicate will consist of a 1-L glass jar containing ~4 cm of sediment and ~800 mL of clean overlying seawater at ~30 ppt. The test conditions include exposure at 20 ± 1°C under continuous light. The tests will be initiated with the random allocation of 20 randomly-selected test organisms into each replicate. Water quality parameters, including pH, temperature, dissolved oxygen (D.O.), and salinity, will be measured daily during testing.

The tests will be terminated after 10 days exposure. The test endpoint is survival, with the test response for the Site Composite being compared to either a reference sediment (i.e., SF-9) or reference sediment database for determination of potential impairment.

Reference Toxicant Testing – In order to assess the sensitivity of the amphipods used in these tests to toxic stress, a reference toxicant test will be run concurrently with the whole sediment amphipod test. The *Ampelisca* reference toxicant test consists of a 96-hr water-only exposure to cadmium (as $CdCl_2$). The test response data for cadmium are then compared to the ongoing database of response data from previous reference toxicant tests performed by the laboratory.

5.2.1.2 Polychaete Solid-Phase Survival Bioassay The second benthic test species will be the marine polychaete *Neanthes arenacoedentata*, obtained from an ongoing culture maintained by Dr. Donald Reish at Long Beach State University. Control sediment will also be collected from a site free from contamination and of known quality to produce acceptable survival.

Polychaete tests will be conducted as 10-day (acute) static exposures, with 5 replicates per treatment. Each replicate will consist of a 1-L glass beaker containing ~2.5 cm of sediment and ~800 mL of clean overlying seawater at ~30 ppt. The test conditions include exposure at 20 ± 1°C under a 12L:12D photoperiod. The tests will be initiated with the random allocation of 10 randomly selected test organisms into each replicate. Water quality parameters, including pH, temperature, D.O., and salinity, will be measured daily during testing.

The tests will be terminated after 10 days exposure. The test endpoint is survival, with the test response for the Site Composite being compared to either a reference sediment (i.e., SF-9) or reference sediment database for determination of potential impairment for determination of potential impairment.

Reference Toxicant Testing – In order to assess the sensitivity of the polychaetes used in these tests to toxic stress, a reference toxicant test will be run concurrently with the whole sediment polychaete test. The *Neanthes* reference toxicant test consists of a 96-hr water-only exposure test using cadmium (as $CdCl_2$). The test response data for cadmium are then compared to the ongoing database of response data from previous reference toxicant tests performed by the lab.

5.2.1.3 Statistical Analyses for the Benthic Sediment Toxicity Tests – The Control treatment acceptability criteria for survival is $\geq$90% survival in the "Home" sediment treatment for both amphipods and polychaetes. The test organism survival data will be analyzed to determine if there are any statistically significant reductions in survival in the DU-1 (Area 1) sediments relative to the appropriate control treatments. All statistical analyses will be performed using CETIS® (TidePool Scientific, McKinleyville, CA). A toxicologically significant effect in the sediment bioassays is defined as a statistically significant reduction in survival and a >20% reduction in survival for amphipods or >10% reduction in survival for polychaetes, relative to their respective reference site treatments or reference site database value.

5.2.2 Sediment Elutriate Water Column Toxicity Testing

Dredged material disposal regulations for ocean disposal require water-column evaluations of the sediment elutriate using species from different phyla where possible. Sediment elutriate tests will be performed using bivalve (*Mytilus galloprovinciales*) embryos as described in ASTM method E724-98, mysid shrimp (*Americamysis bahia*) as described in EPA/821/R-02/012 (Test Method 2007.0), and the inland silverside (*Menidia beryllina*) as described in EPA/821/R-02/012 (Test Method 2006.0).

Elutriate toxicity samples will be prepared as per ITM/OTM procedures, mixing a slurry of 1 part sediment to 4 parts Site Water for 30 minutes at room temperature (~22°C), followed by a 60-minute settling period (post-settling centrifugation may be implemented, if necessary to remove suspended fines). The resulting supernatant is considered 100% elutriate. If the salinity of the site water is ≤28 ppt, the site water will either be adjusted up to a salinity of 30±2 ppt via addition of artificial sea salts prior to use, or clean seawater collected from the UC Davis Granite Canyon Marine Laboratory (Carmel, CA) will be diluted to a salinity of 30±2 ppt via addition of reverse-osmosis- de-ionized water for use in the elutriate preparation.

5.2.2.1 Water Column *Mytilus galloprovinciales* Embryo-Larval Development Bioassay – The Control water for these tests will consist of 0.45-μm-filtered clean seawater (from the UC Davis Granite Canyon Marine Laboratory), diluted to ~30 ppt salinity via addition of reverse-osmosis, de-ionized water. The 100% elutriate and the Control water will be used to prepare test solutions at concentrations of 1%, 10%, and 50% elutriate. If ammonia concentrations in the bulk sediment exceed 15 mg/L total ammonia-N, an additional 25% elutriate concentration may be included; this additional concentration has proven useful in the past in differentiating between chemical-related effects and ammonia-related effects. Routine water quality characteristics will be determined for each test solution prior to use in these tests.

There will be 5 replicates for each treatment, each replicate consisting of 10-mL of test solution within a 20-mL glass scintillation vial. The tests will be initiated by the random allocation of 150-300 embryos into each test replicate, which will then be placed into a temperature-controlled water bath at 16°C under a 16L:8D photoperiod.

After 48 (±2) hrs exposure, the tests will be terminated, and the contents of each test replicate vial will be preserved via addition of 5% glutaraldehyde. The preserved embryos will be examined microscopically to determine the percentage survival and percentage normal embryo development of the test organisms. The resulting survival and embryo development data are then statistically analyzed and key dose-response LC and EC point estimates determined for each site sediment elutriate using the CETIS® statistical software.

Reference Toxicant Testing – In order to assess the sensitivity of the *Mytilus* embryos used in these tests to toxicant stress, a reference toxicant test will be performed. The reference toxicant test will be performed similarly to the sediment elutriate tests, but will use test solutions

consisting of Lab Control water spiked with copper (as $CuSO_4$), at concentrations of 1.25, 2.5, 5, 10, 15, and 20 μg/L instead of elutriate dilutions. The resulting test response data will be analyzed to determine key dose-response point estimates (e.g., EC_{50}); all statistical analyses will be made using the CETIS® software. These response endpoints will then be compared to the typical response range established by the mean ± 2 SD of the point estimates generated by the 20 most recent previous reference toxicant tests performed by this lab.

5.2.2.2 Water Column *Americamysis bahia* Acute Toxicity Test - The Control water for these tests will consist of 0.45-μm-filtered clean seawater (from the UC Davis Granite Canyon Marine Laboratory), diluted to ~30 ppt salinity via addition of reverse-osmosis, de-ionized water. The 100% elutriate and the Control water will be used to prepare test solutions at concentrations of 1%, 10%, and 50% elutriate. If ammonia concentrations in the bulk sediment exceed 15 mg/L total ammonia-N, an additional 25% elutriate concentration may be included; this additional concentration has proven useful in the past in differentiating between chemical-related effects and ammonia-related effects. Routine water quality characteristics will be determined for each test solution prior to use in these tests.

There will be 5 replicates for each treatment, each replicate consisting of 200-mL of test solution within a 600-mL beaker. The tests will be initiated by the random allocation of 10 mysids into each test replicate, which will then be placed into a temperature-controlled room at 20°C under a 16L:8D photoperiod.

Each day, water quality conditions will be determined for one randomly-selected replicate per treatment, and the test replicates are examined to determine the number of surviving organisms, with any dead organisms being removed via pipette. After ~48 hrs, each replicate is fed brine shrimp nauplii.

After 96 (±2) hrs exposure, the tests are terminated. At test termination, the final water quality conditions are determined for one randomly-selected replicate per treatment, after which each of the test replicates will be examined to determine the number of surviving mysids. The resulting survival data will then be statistically analyzed and key dose-response EC point estimates determined for each site sediment elutriate using the CETIS® statistical software.

Reference Toxicant Testing – In order to assess the sensitivity of the test organisms to toxic stress, a reference toxicant test is performed concurrently with the elutriate tests. The reference toxicant test is performed similarly to the sediment elutriate tests, but uses test solutions consisting of artificial seawater (reverse-osmosis, de-ionized water adjusted to a salinity of 25 ppt using an artificial sea salt [Crystal Seas® – bioassay grade]) spiked with chromium (as $K_2Cr_2O_7$) at test concentrations of 0.88, 1.75, 3.5, 7, 14, and 28 mg/L, instead of elutriate dilutions. The resulting survival data are statistically analyzed to generate key dose-response EC point estimates; all statistical analyses are performed using the CETIS® statistical package. The results of this test will then be compared to our database of performance by these organisms in

previous reference toxicant tests to determine if the current response is consistent with historical results (i.e., within the range established by the historical mean ± 2 S.D.).

5.2.2.3 Water Column *Menidia beryllina* Acute Toxicity Test – The Control water for these tests will consist of 0.45-μm-filtered clean seawater (from the UC Davis Granite Canyon Marine Laboratory), diluted to ~30 ppt salinity via addition of reverse-osmosis, de-ionized water. The 100% elutriate and the Control water will be used to prepare test solutions at concentrations of 1%, 10%, and 50% elutriate. If ammonia concentrations in the bulk sediment exceed 15 mg/L total ammonia-N, an additional 25% elutriate concentration may be included; this additional concentration has proven useful in the past in differentiating between chemical-related effects and ammonia-related effects. Routine water quality characteristics will be determined for each test solution prior to use in these tests.

There will be 5 replicates for each treatment, each replicate consisting of 200-mL of test solution within a 600-mL beaker. The tests will be initiated by the random allocation of 10 fish into each test replicate, which will then be placed into a temperature-controlled room at 20°C under a 16L:8D photoperiod.

Each day, water quality conditions will be determined for one randomly-selected replicate per treatment, and the test replicates are examined to determine the number of surviving organisms, with any dead organisms being removed via pipette. After ~48 hrs, each replicate is fed brine shrimp nauplii.

After 96 (±2) hrs exposure, the tests are terminated. At test termination, the final water quality conditions are determined for one randomly-selected replicate per treatment, after which each of the test replicates will be examined to determine the number of surviving fish. The resulting survival data will then be statistically analyzed and key dose-response EC point estimates determined for each site sediment elutriate using the CETIS® statistical software.

Reference Toxicant Testing – In order to assess the sensitivity of the test organisms to toxic stress, a reference toxicant test is performed concurrently with the elutriate tests. The reference toxicant test is performed similarly to the sediment elutriate tests, but used test solutions consisting of artificial seawater (reverse-osmosis, de-ionized water adjusted to a salinity of 25 ppt using an artificial sea salt [Crystal Seas® – bioassay grade]) spiked with copper (as Cu_2SO_4) at test concentrations of 32, 64, 128, 256, and 512 μg/L, instead of elutriate dilutions. The resulting survival data are statistically analyzed to generate dose-response key LC point estimates; all statistical analyses are performed using the CETIS® statistical package. The results of this test will then be compared to our database of performance by these organisms in previous reference toxicant tests to determine if the current response is consistent with historical results (i.e., within the range established by the historical mean ± 2 S.D.).

5.2.2.4 Statistical Analyses for the Water Column Toxicity Tests – The test acceptability criteria for Control treatment are ≥70% survival and ≥70% normal development for bivalve tests and ≥90% survival for the mysid and fish tests. Key point estimates (e.g., LC_{50} and EC_{50} values) will be determined for the elutriate tests following the EPA statistical analysis flowchart. All statistical analyses will be performed using CETIS®.

5.2.3 Modified Elutriate Test (MET) Toxicity Testing

As per the SFRWQCB WDR order #R2-2005-0034 for the Hamilton Wetland Restoration Project, a toxicity test will be performed on the resulting MET elutriate; testing will be performed using mysid shrimp (*Americamysis bahia*) as described in EPA/821/R-02/012 (Test Method 2007.0).

Elutriate toxicity samples will be prepared as per procedures described in USACE 1985 and USACE 2003, by mixing a slurry of 150 g/L of sediment (dry wt) in Site Water for 5 minutes at room temperature (~22°C), aeration for 1 hour, followed by a 24 hr settling period. The resulting supernatant is considered 100% 'modified' elutriate. If the salinity of the site water is ≤20 ppt, the site water will either be adjusted up to a salinity of 20±2 ppt via addition of artificial sea salts prior to use, or clean seawater collected from the UC Davis Granite Canyon Marine Laboratory (Carmel, CA) will be diluted to a salinity of 20±2 ppt via addition of reverse-osmosis- de-ionized water for use in the elutriate preparation.

5.2.4 Benthic Sediment Bioaccumulation Testing

Bioaccumulation tests are designed to evaluate the potential of benthic organisms to accumulate contaminants from contaminated sediment. Bioaccumulation tests are based on analysis of the organisms' tissues after 10 or 28 days of exposure. The 10-day exposure test is appropriate when the only contaminants of concern are metals; 28-day tests should be used when any contaminants of concern are organic or organometallic.

The two species proposed for benthic bioaccumulation testing are the bivalve, *Macoma nasuta* and the polychaete, *Nephtys caecoides*. These tests will be performed using ASTM method E1688-97a.

5.2.4.1 Solid-Phase Bioaccumulation Bioassay with the Bivalve *Macoma nasuta* – The first benthic bioaccumulation test species will be the marine bivalve *Macoma nasuta*. Control sediment will collected from a site free from contamination and of known quality to produce acceptable survival.

There will be 5 replicates for each treatment, each replicate consisting of 4 L of sediment placed within a 10 L HD polyethylene tank. Clean seawater (1 μm-filtered seawater from the UC Davis Granite Canyon Marine Laboratory) is carefully poured into each tank so as to minimize disturbance of the sediment. The replicate tanks are then placed into a temperature controlled room under aeration at 12°C.

After 24 hrs equilibration, routine water quality characteristics (pH, D.O., and salinity) are determined for each test replicate at each treatment. Then, 20-25 randomly-selected adult clams are placed into each replicate container. Additional bivalves are also transferred to clean sand (to promote depuration) at this time for determination of T_0 tissue concentrations (these tissues will be harvested after 24 hrs, and the tissues processed and frozen for later analyses, as described below). Each day, for the prescribed test duration, the D.O. of the overlying water is measured in one test replicate for each treatment. Approximately 80% of the overlying water in each replicate is carefully replaced three times per week; immediately after each water change, the D.O. and salinity are measured in one test replicate for each treatment.

After the prescribed test duration, the bivalves are transferred into clean containers containing clean sand to allow the organisms to depurate the test sediment. After this purging process, the organisms are rinsed with clean seawater and the shell length is then measured to the nearest mm. The organisms are then placed into an appropriate size container, and immediately frozen. The frozen clams are then shipped to the appropriate analytical laboratories for analysis of potential contaminants.

Upon arrival at the analytical laboratory, the soft tissue contents of each bivalve are removed using stainless steel forceps and scalpel, rinsed with de-ionized water and blot-dried, and then weighed to the nearest 0.1 gm. The soft tissue samples from each replicate treatment are composited, homogenized in a stainless steel blender, and placed into pre-cleaned glass vials, which are sealed and labeled for identification and subsequent analysis.

5.2.4.2 Solid-Phase Bioaccumulation Bioassay with the Polychaete *Nephtys caecoides* – The second benthic bioaccumulation test species will be the marine polychaete *Nephtys caecoides*. Control sediment will also be collected from a site free from contamination and of known quality to produce acceptable survival.

There will be 5 replicates for each treatment, each replicate consisting of 4 L of sediment placed within a 10 L HD polyethylene tank. Clean seawater (1 um-filtered seawater from the UC Davis Granite Canyon Marine Laboratory) is carefully poured into each tank so as to minimize disturbance of the sediment. The replicate tanks are then placed into a temperature controlled room under aeration at 12°C.

After 24 hrs equilibration, routine water quality characteristics (pH, D.O., and salinity) are determined for each test replicate at each treatment. Then, 50 randomly-selected polychaetes are placed into each replicate container. Additional polychaetes are also transferred to clean sand (to promote depuration) at this time for determination of T_0 tissue concentrations (these tissues are harvested after 24 hrs, and the tissues processed and frozen for later analyses, as described below). Each day, for the prescribed test duration, the D.O. of the overlying water is measured in one test replicate for each treatment. Approximately 80% of the overlying water in each replicate

is carefully replaced three times per week; immediately after each water change, the D.O. and salinity are measured in one test replicate for each treatment.

After the prescribed test duration, the polychaetes are sieved from the sediment, and enumerated to determine the number of surviving organisms (for potential use as an assessment of toxicity), and then transferred into clean containers containing clean sand to allow the organisms to depurate the test sediment. After this purging process, the organisms are rinsed with clean seawater and then placed into an appropriate size container, and immediately frozen. The frozen polychaetes are then shipped to the appropriate analytical laboratories for analysis of potential contaminants.

Upon arrival at the analytical laboratory, each polychaete is removed using stainless steel forceps and scalpel, rinsed with de-ionized water and blot-dried, and then weighed to the nearest 0.1 gm. The tissue samples from each replicate treatment are composited, homogenized in a stainless steel blender, and placed into pre-cleaned glass vials, which are sealed and labeled for identification and subsequent analysis.

5.3 Quality Assurance (QA) Objectives

Quality assurance procedures to be used for sediment characterization and testing are consistent with methods described in USEPA/USACE (1991, 1995, 1998) and USEPA (1998, 2002). The methods employed in this sediment sampling and characterization program are detailed in standard guides (*e.g.*, Standard Methods, ASTM, USEPA, etc.) and Standard Operating Procedures are maintained in the bioassay and analytical laboratories.

All QA/QC records for the various testing programs are kept on file for review by regulatory personnel.

5.3.1 Chemical and Physical Analyses Quality Assurance

5.3.1.1 Accuracy - Accuracy estimates will be based on analyses of lab blanks, analytical recoveries of matrix spikes of test samples and laboratory control materials, and analysis of certified reference material. Results from spikes and/or reference materials are reported as "percent recovery", determined by comparing the measured analyte concentrations of the Standard Reference Materials, Laboratory Control Materials, or matrix spikes to the "True Value". Percent Recovery will be reported along with the corresponding acceptance ranges. Where possible, surrogate compounds will be spiked into each sample and surrogate percent recovery will be reported along with the corresponding control limits.

Matrix spikes are added prior to processing the sample and carried through the entire analytical procedure. Matrix spike data for both trace metals and organics will be provided at a frequency of one set of duplicate spikes per QA batch.

5.3.1.2 Precision - Precision will be estimated by analyzing duplicate samples and matrix spike duplicate samples. Duplicate analyses are performed on actual site samples and not on reference site samples. Results from duplicate analyses of the actual test samples may also indicate homogeneity of the sample matrix. Relative percent differences (RPDs) are calculated for all duplicate samples or spikes and are reported along with acceptance ranges (typically 0-30%).

5.3.1.3 Analytical Methods - All sample analyses will be performed using EPA Methods, where applicable (see above for method specification for each analyte group). Daily logs of instrument performance are maintained, including initial and continuing calibration verification.

5.3.2 Biological Testing Quality Assurance

All sediment toxicity tests will incorporate standard toxicity testing QA/QC procedures to ensure that the test results are valid. Standard QA/QC procedures include the use of negative controls, positive controls (reference toxicant tests), reference sediment samples, replicates, and measurements of water quality during testing.

5.3.2.1 Water and Sediment Handling and Storage - Sediment samples will be maintained at 4°C in the dark until they are used in the bioassay testing system. All sediments are held in sealed, labeled sample storage bags. Site water samples will be similarly stored in sealed, labeled containers at 4°C. Seawater used in these tests will come from the UC Davis Granite Canyon Marine Laboratory (Carmel, CA), and will be stored on-site at PER in an insulated 3,000 gallon HDPE tank at 4°C. Sub-samples designated for long-term storage are archived under the appropriate holding conditions.

5.3.2.2 Source and Condition of Test Organisms - All test organisms will be obtained from reputable suppliers who have provided PER with organisms in the past. Normally, all test organisms are maintained in the laboratory for acclimation to test conditions (exceptions are bivalves). If mortality in excess of 5% is noted in the holding stock, the animals will be discarded and a new batch ordered.

5.3.2.3 Maintenance of Test Conditions and Corrective Actions - Each of the biological tests has a set of specific test conditions that are defined in the standard testing. For example, water quality measurements will be monitored to ensure that test conditions are within the prescribed limits for each test procedure. The limits for various test condition parameters are noted in the section on the acceptability of each test. If these criteria are not met, the test may be re-run if appropriate.

5.3.2.4 Calibration Procedures and Frequency - Instruments are calibrated daily according to Laboratory (SOPs) and calibration data are logged and initialed. Calibration logs are monitored weekly to ensure completeness.

5.3.2.5 Reference Toxicant Testing and Data Accuracy and Precision - The accuracy of toxicity tests (e.g., LC_{50} point estimates) are not normally measured in biological testing. Instead, concurrent reference toxicant tests are used to assess accuracy and precision. For instance, acceptable accuracy is defined as a current measured LC_{50} reference toxicant value that is within 2 standard deviations of the current laboratory mean established by previously performed reference toxicant tests. A reference toxicant will be performed concurrently with the testing for each species to establish that the test organisms are responding to toxic stress in a typical fashion.

The precision of toxicity tests is assessed via measures of variability (e.g., coefficient of variation [CV] for a given test treatment). While there are no "acceptability limits" placed on the CV for most test responses, these can be evaluated using "Best Professional Judgment" to characterize whether or not the test response at a given treatment is subject to too much variability for use in a given test.

5.3.2.6 Data Evaluations - Bioassay tests are performed according to accepted protocols and standard test conditions. All test data, data analyses, and other relevant records for each test will be reviewed for accuracy and completeness by the quality control unit. Deviations from the standard testing guides are reported with the final report. If and when such deviations are observed, the test will be evaluated to determine whether it is valid according to the regulatory agency to which it will be submitted. If it is determined to be invalid, the client will be notified and the test rerun.

5.3.2.7 Sample Tracking - Sample COC sheets, sample receipt logs, sample holding, and sample labeling procedures are audited weekly by the quality control unit. Sub-samples designated for long-term storage are archived under the appropriate holding conditions.

5.3.3 Deviations from Protocol

Any deviations from approved SOP's or this SAP will be summarized and qualified with respect to how they may have affected data quality.

6. DATA MANAGEMENT

Analytical results will be provided by all subcontract analytical laboratories in both hard copy and electronic format. All data will be reviewed by the PER Project Manager to ensure that the data quality objectives for each analysis are met and that both the electronic and hard copy forms of data are accurate. Hard copies of all data reports will be placed in the project files at PER; electronic data reports will be archived on PER's server, and will be available for electronic transfer to marina staff and the DMMO, if requested.

7. DATA ANALYSIS AND INTERPRETATION

Data will be analyzed and presented clearly so that suitability for disposal at an unconfined aquatic disposal or an upland wetland reuse site can be determined. All analytical data will be reviewed for accuracy prior to reporting; data will be presented in tabular form. The physical and chemical characteristics of sediment samples will be evaluated according to the DMMO review process; if the results indicated that the material would be suitable for unconfined aquatic disposal then biological testing will be performed. If performed, benthic sediment toxicity test results will be compared to the SF-11 and SF-DODS reference database according to the DMMO review process; water column toxicity test results will be compared to Elutriate Suitability Concentrations (ESC) at the edge of the mixing zone for the SF-11 and SF-DODS Disposal Sites. Bioaccumulation test results will be compared to the SF-DODS reference database.

7.1 Sediment Chemistry and Conventional Data Analyses

Sediment physical and chemical characteristics provide information about chemicals of concern present in the sediment and their potential bioavailability, and about non-chemical factors that could affect toxicity. Data analysis of sediment chemistry and conventional parameters will consist of tabulation and comparison with existing regulatory guidelines (USEPA/USACE 1991, 1998) as requested by the DMMO. Sediment chemistry results will also be used to identify "hot spots" which may need further resolution (e.g., analysis of sediment material from individual cores), and/or to assist in evaluating appropriate disposal options).

7.2 Benthic Toxicity Test Data

ITM and OTM guidance requires that test sediment results be compared with disposal site and/or reference site sediment results or a reference site database (SF-DODS) to determine the potential impact of whole sediment on benthic organisms at and beyond the boundaries of the disposal site (USEPA/USACE 1998). As detailed in the ITM and OTM, comparative guidelines for acceptance are listed below:

1. If survival is greater in the proposed dredged sediments than in reference site sediment(s) or the reference site sediment database, the proposed dredged sediments are <u>not</u> acutely toxic to benthic organisms.
2. If there is $\leq$20% reduction in amphipod survival in the site sediment relative to the reference sediment survival (or the 'reference site database survival'), the test sediments are <u>not</u> acutely toxic to the amphipods. If there is >20% reduction in survival between a test sediment and the reference sediment, then the survival response for the test sediment must be compared statistically to the reference sediment; if the difference in survival is statistically significant, then the test sediments are considered to be acutely toxic to the amphipods (statistical analyses are not performed when reference site database values are used).

3. If there is ≤10% reduction in polychaete survival, in the site sediment relative to the reference sediment survival (or the 'reference site database survival'), the test sediments are not acutely toxic to the polychaetes. If there is >10% reduction in survival between a test sediment and the reference sediment, then the survival response for the test sediment must be compared statistically to the reference sediment; if the difference in survival is statistically significant, then the test sediments are considered to be acutely toxic to the polychaetes (statistical analyses are not performed when reference site database values are used).

7.3 Water Column (Sediment Elutriate or Liquid Suspended Phase) Toxicity Test Data

Comparative guidelines for interpretation of water column tests, as detailed in the ITM ad OTM, are listed below:

1. If survival and normal embryo development in the 100% sediment elutriate treatment is ≥ than survival in the Control (clean seawater) treatment, the dredged material is not predicted to be acutely toxic to water column organisms.
2. If there is ≤10% reduction in survival or normal embryo development in a 100% sediment elutriate relative to the Control treatment response, there is no need for statistical analyses and no indication of water column toxicity attributable to the test sediments.
3. If there is >10% reduction in survival or normal embryo development in the 100% sediment elutriate relative to the Control treatment response, then data must be evaluated statistically to determine the magnitude of toxicity. If there is >50% survival or normal embryo development in the 100% elutriate treatment, the LC_{50}/EC_{50} is assumed to be ≥100%. If there is <50% survival or normal embryo development in at least one of the elutriate treatments, then an LC_{50}/EC_{50} should be calculated and compared with existing acceptability standards.

7.3.1 Dilution Model Calculations

The Short Term Fate Model (STFATE) for open water barge and hopper discharges will be used to model the fate of disposed sediments and determine if water quality criteria will be met at the edge of the mixing zone for the disposal site; input parameters, unique to the SF-DODS site, will be used. A sample will exceed water quality criteria if 1% of the calculated LC_{50} or EC_{50} (whichever is more conservative) is lower than the projected suspended phase concentration of the dredge material at the edge of the mixing zone. Based on the results of the water column toxicity tests, modeling with STFATE may not be required. A generic model, approved by the DMMO, will be used to determine the sediment concentration at the edge of the mixing zone with respect to the SF-11 disposal site.

8. REPORTING AND DELIVERABLES

8.1 Sampling and Analysis Results

PER will prepare a Final Sampling & Analysis Data Report documenting all activities associated with the collection, transportation, handling (e.g. compositing), sample shipment, and chemical and conventional analyses, and biological testing of the sediment samples. All Lab Data Reports received from sub-contracting analytical laboratories will be included as Appendices to the Final Data Report. At a minimum, the following will be included in the Final Data Report:

1. Summary of all field activities, including a description of any deviations from the approved SAP;
2. Locations of sediment sampling stations in latitude and longitude (in degrees and minutes to 3 decimal places). All vertical elevations of mud-line and water surface will be reported to the nearest 0.1 ft relative to MLLW;
3. A project map with actual sampling locations;
4. Analytical data results and QA/QC review; and,
5. Summary of comparison of chemical results.

9. REFERENCES

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PER 2006b. Characterization of Levin Richmond Terminal Site LRT-S01 Sediment Core Samples for Total DDT. Prepared for Cooper White & Cooper, Walnut Creek, CA 94596. Prepared by Pacific EcoRisk, Martinez, CA 94553.

PER 2006c. Characterization of Shore Terminal Sediments: Results of Dredge Materials Sampling and Analysis. Prepared for Cooper White & Cooper, Walnut Creek, CA 94596. Prepared by Pacific EcoRisk, Martinez, CA 94553.

PER 2006d. Characterization of Shore Terminal Site LRT-S02 Sediment Core Samples for Total DDT. Prepared for Cooper White & Cooper, Walnut Creek, CA 94596. Prepared by Pacific EcoRisk, Martinez, CA 94553.

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Appendix A

Recent Testing History

Levin-Richmond Terminal Berth A

Figure A-1. Site LRT-S01 (Levin-Richmond Terminal) Sediment Core Locations

Table A-1. Results of grain size analyses of Levin-Richmond sediments

Analytes	LRT-SO1 COMP	Method Reporting Limit
% Gravel	3.30	0.1
% Sand	18.2	0.1
% Silt	30.1	0.1
% Clay	46.1	0.1

Table A-2. Results of conventional analyses of Levin-Richmond sediments

Analytes	LRT-SO1 COMP	Method Reporting Limit
Total Solids (% as Dry Wt.)	44.4	0.1
Total Organic Carbon (%)	1.72	0.1

Table A-3. Metals concentrations (mg/kg, dry wt.) of Levin-Richmond sediments

Metals	LRT-SO1 COMP	Method Reporting Limit
Arsenic	7.1	0.5
Cadmium	0.44	0.05
Chromium	78.9	1.0
Copper	48.1	0.1
Lead	35.2	0.05
Mercury	0.31	0.02
Nickel	56.6	0.2
Selenium	0.2	0.1
Silver	0.32	0.02
Zinc	95.3	0.5

Table A-4. PAH concentrations (μg/kg, dry wt) of Levin-Richmond sediments

PAHs	LRT-SO1 COMP	Method Reporting Limit
Acenaphthene	26	5.7
Acenaphthylene	46	5.7
Anthracene	160	5.7
Benzo(a)anthracene	350	5.7
Benzo(a)pyrene	530	5.7
Benzo(b)fluoranthene	510	5.7
Benzo(g,h,i)perylene	220	5.7
Benzo(k)fluoranthene	390	5.7
Chrysene	740	5.7
Dibenzo(a,h)anthracene	74	5.7
Dibenzofuran	16	5.7
Fluoranthene	430	5.7
Fluorene	30	5.7
Indeno(1,2,3-cd)pyrene	220	5.7
Methylnaphtalene	18	5.7
Naphthalene	34	5.7
Phenanthrene	140	5.7
Pyrene	730	5.7
Total PAHs	4664	NA

Table A-5. Organochlorine pesticide concentrations (μg/kg, dry wt.) of Levin-Richmond sediments

Organochlorine Pesticides	LRT-SO1 COMP	Method Reporting Limit
Aldrin	<1	1
a-BHC	<1	1
b-BHC	<1.0	1.1
g-BHC (Lindane)	<1	1
d-BHC	<1	1
alpha-Chlordane	<1	1
gamma-Chlordane	<1.7	1.7
Dieldrin	8.7	1
Endosulfan I	<1	1
Endosulfan II	1.3	1
Endosulfan sulfate	<1	1
Endrin	<1	1
Endrin aldehyde	<1	1
Endrin ketone	<1	1
Heptachlor	<1	1
Heptachlor epoxide	1.7	1
Methoxychlor	<1	1
Toxaphene	<84	84
4,4'-DDD	<1	10
4,4'-DDE	28	1
4,4'-DDT	<1	1
Total DDT	28	NA

Table A-6. Total DDT concentrations (μg/kg, dry wt.) of Levin Richmond LRT-S01 individual sediment core samples

Analyte	LRT-S01-01	LRT-S01-02	LRT-S01-03	LRT-S01-04	LRT-S01-05	Method Reporting Limit
4,4'-DDD	170	190	170	170	260	18
4,4'-DDE	49	52	45	30	51	18
4,4'-DDT	86	220	160	74	73	18
Total DDT	305	462	375	274	384	NA

Table A-7. Organotin concentrations (μg/kg, dry wt.) of Levin-Richmond sediments

Organotins	LRT-SO1 COMP	Method Reporting Limit
Monobutyltin	2.7	2.3
Dibutyltin	13	2.3
Tributyltin	30	2.3
Tetrabutyltin	<2.3	2.3
Total Butyltins	45.7	NA

Table A-8. PCB Aroclor concentrations (μg/kg, dry wt) of Levin-Richmond sediments

PCB Aroclors	LRT-SO1 COMP	Method Reporting Limit
Aroclor 1016	<10	10
Aroclor 1221	<20	20
Aroclor 1232	<10	10
Aroclor 1242	<10	10
Aroclor 1248	<10	10
Aroclor 1254	<79	79
Aroclor 1260	<10	10
Total PCBs	<10	NA

Table A-9. *Ampelisca abdita* survival in the solid-phase test sediments

Sediment Site	% Survival in Test Replicates					Overall Mean % Survival
	Rep A	Rep B	Rep C	Rep D	Rep E	
"Home" Lab Control	100	95	95	90	90	**94**
Alcatraz (SF-11)	70	75	80	80	75	**76**
San Pablo (SF-10)	75	100	65	85	65	**78**
LRT-SO1 COMP	95	90	95	75	90	**89**

Table A-10. *Neanthes arenaceodentata* survival in the test sediments

Sediment Site	% Survival in Test Replicates					Overall Mean % Survival
	Rep A	Rep B	Rep C	Rep D	Rep E	
"Home" Lab Control	100	90	100	100	100	**98**
Alcatraz (SF-11)	100	100	100	100	100	**100**
San Pablo (SF-10)	100	100	100	100	100	**100**
LRT-SO1 COMP	100	100	100	100	90	**98**

Table A-11. Effects of LRT-SO1 COMP sediment elutriate on *Mytilus sp.* embryos

Elutriate Treatment	Mean % Survival	Mean % Normal Development
Lab Control	93	93
1%	93	90
10%	89	93
25%	95	94
50%	87	86
100%	0	0
LC_{50} or EC_{50} =	66.6% elutriate	73.1% elutriate
Disposal limit met?	Yes	Yes

Pacific Atlantic (formally Shore) Terminal

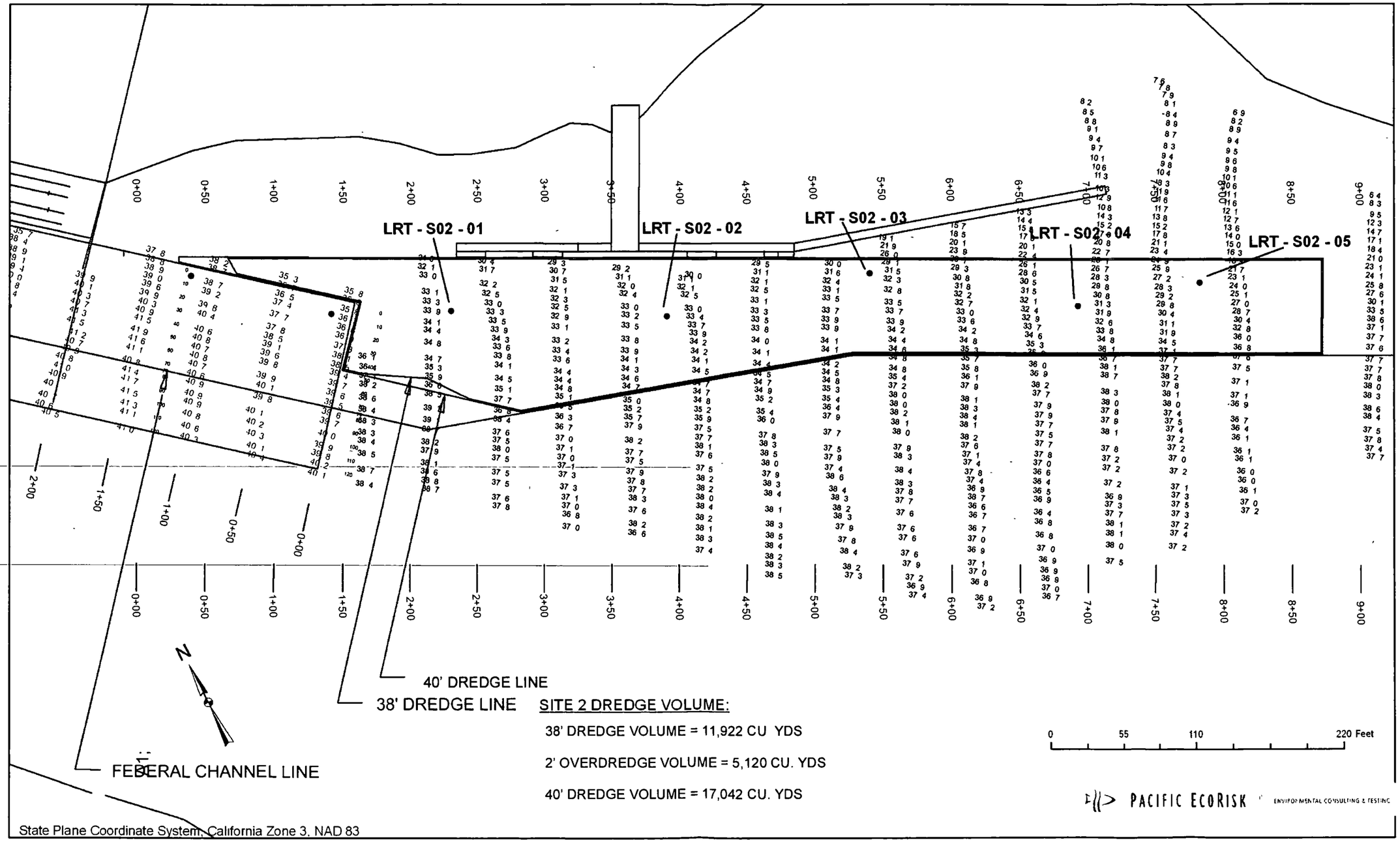

Figure A-2. Site LRT-S02 (Shore Terminal) Sediment Core Locations

Table A-12. Results of grain size analyses of Shore Terminal sediments

Analytes	LRT-SO2 COMP	Method Reporting Limit
% Gravel	0.00	0.1
% Sand	13.3	0.1
% Silt	38.5	0.1
% Clay	49.6	0.1

Table A-13. Results of conventional analyses of Shore Terminal sediments

Analytes	LRT-SO2 COMP	Method Reporting Limit
Total Solids (% as Dry Wt.)	44.7	0.1
Total Organic Carbon (%)	1.14	0.1

Table A-14. Metals concentrations (mg/kg, dry wt.) of Shore Terminal sediments

Metals	LRT-SO2 COMP	Method Reporting Limit
Arsenic	7.0	0.5
Cadmium	0.40	0.05
Chromium	83.0	1.0
Copper	39.3	0.1
Lead	30.1	0.05
Mercury	0.35	0.02
Nickel	59.7	0.2
Selenium	0.2	0.1
Silver	0.38	0.02
Zinc	82.8	0.5

Table A-15. PAH concentrations (μg/kg, dry wt) of Shore Terminal sediments

PAHs	LRT-SO2 COMP	Method Reporting Limit
Acenaphthene	<1	5.6-5.7
Acenaphthylene	<1	5.6-5.7
Anthracene	<1	5.6-5.7
Benzo(a)anthracene	8.1	5.6-5.7
Benzo(a)pyrene	11	5.6-5.7
Benzo(b)fluoranthene	12	5.6-5.7
Benzo(g,h,i)perylene	12	5.6-5.7
Benzo(k)fluoranthene	9.3	5.6-5.7
Chrysene	12	5.6-5.7
Dibenzo(a,h)anthracene	<1	5.6-5.7
Dibenzofuran	<1	5.6-5.7
Fluoranthene	14	5.6-5.7
Fluorene	<1	5.6-5.7
Indeno(1,2,3-cd)pyrene	10	5.6-5.7
Methylnaphtalene	<1	5.6-5.7
Naphthalene	<1	5.6-5.7
Phenanthrene	6.0	5.6-5.7
Pyrene	16	5.6-5.7
Total PAHs	110.4	NA

Table A-16. Organochlorine pesticide concentrations (μg/kg, dry wt.) of Shore Terminal sediments

Organochlorine Pesticides	LRT-SO2 COMP	Method Reporting Limit
Aldrin	<1	1
a-BHC	<1	1
b-BHC	<1.1	1.1
g-BHC (Lindane)	<1	1
d-BHC	<1	1
alpha-Chlordane	<1	1
gamma-Chlordane	<1.6	1.6
Dieldrin	3.4	1
Endosulfan I	<1	1
Endosulfan II	3.1	1
Endosulfan sulfate	<1	1
Endrin	<1	1
Endrin aldehyde	<1	1
Endrin ketone	1.4	1
Heptachlor	<1	1
Heptachlor epoxide	1.2	1
Methoxychlor	<1	1
Toxaphene	<50	50
4,4'-DDD	83	10
4,4'-DDE	20	1
4,4'-DDT	35	1
Total DDT	138	NA

Table A-17. Total DDT concentrations (μg/kg, dry wt.) of Shore Terminal LRT-S02 individual sediment core samples

Analyte	LRT-S02-01	LRT-S02-02	LRT-S02-03	LRT-S02-04	LRT-S02-05	Method Reporting Limit
4,4'-DDD	160	180	87	120	85	20
4,4'-DDE	31	35	21	26	33	20
4,4'-DDT	49	75	32	24	36	20
Total DDT	240	290	140	170	154	NA

Table A-18. Organotin concentrations (μg/kg, dry wt.) of Shore Terminal sediments

Organotins	LRT-SO2 COMP	Method Reporting Limit
Monobutyltin	<2.3	2.3
Dibutyltin	11	2.3
Tributyltin	18	2.3
Tetrabutyltin	<2.3	2.3
Total Butyltins	29	NA

Table A-19. PCB Aroclor concentrations (μg/kg, dry wt) of Shore Terminal sediments

PCB Aroclors	LRT-SO2 COMP	Method Reporting Limit
Aroclor 1016	<10	10
Aroclor 1221	<20	20
Aroclor 1232	<10	10
Aroclor 1242	<10	10
Aroclor 1248	<10	10
Aroclor 1254	<31	31
Aroclor 1260	<10	10
Total PCBs	<10	NA

Table A-20. *Ampelisca abdita* survival in the solid-phase test sediments

Sediment Site	% Survival in Test Replicates					Overall Mean % Survival
	Rep A	Rep B	Rep C	Rep D	Rep E	
"Home" Lab Control	100	95	95	90	90	**94**
Alcatraz (SF-11)	70	75	80	80	75	**76**
San Pablo (SF-10)	75	100	65	85	65	**78**
LRT-SO2 COMP	85	85	85	80	80	**83**

Table A-21. *Neanthes arenaceodentata* survival in the solid-phase test sediments

Sediment Site	% Survival in Test Replicates					Overall Mean % Survival
	Rep A	Rep B	Rep C	Rep D	Rep E	
"Home" Lab Control	100	90	100	100	100	**98**
Alcatraz (SF-11)	100	100	100	100	100	**100**
San Pablo (SF-10)	100	100	100	100	100	**100**
LRT-SO2 COMP	100	100	90	90	100	**96**

Table A-22. Effects of LRT-SO2 COMP sediment elutriate on *Mytilus sp.* embryo survival & development

Elutriate Treatment	Mean % Survival	Mean % Normal Development
Lab Control	93	93
1%	64	59
10%	91	83
25%	81	80
50%	66	62
100%	0	0
LC_{50} or EC_{50} =	57.9% elutriate	62.4% elutriate
Disposal limit met?	Yes	Yes

Appendix B

Sample Containers, Holding Time, Preservation, and Storage for Analytical Chemistry

Sample Containers, Holding Times, Preservation and Storage

Parameter	Container Type/Size	Holding Time[a]	Preservation/Storage
Metals[b]	125-mL glass jar	Mercury – 28 days All others – 6 months	Hold at 4° ± 2°C up to 1 month or freeze at -20° ± 10°C
Butyltins	500-mL glass with Teflon® lid	14 days to extraction[c]; 40 days to analysis after extraction	Freeze for extended storage (-20° ± 10°C); otherwise store at 4° ± 2°C
PCBs[d], pesticides[e], PAHs[f]	500-mL glass with Teflon® lid	14 days to extraction[c]; 40 days to analysis after extraction	Freeze for extended storage (-20° ± 10°C); otherwise store at 4° ± 2°C
SVOCs TPH-Diesel TPH-Gasoline Phenol Pentachlorophenol MCPA MCPP Dichlorprop Dioxins	500-mL glass with Teflon® lid	14 days to extraction[c]; 30 days to extraction[c] for Dioxins[c]; 40 days to analysis after extraction	Freeze for extended storage (-20° ± 10°C); otherwise store at 4° ± 2°C
Grain size	125-mL plastic	6 months	4° ± 2°C
Total solids, TOC, ammonia	250-mL glass with Teflon® lid	Total solids, TOC – 1 month; ammonia – 7 days	4° ± 2°C
Toxicity tests	4-L glass with Teflon® lid (1 container per acute test)	6 weeks	4° ± 2°C/dark/airtight
Archive	500-mL and 1-L glass jars with Teflon® lid (for composite samples)	1 year	Freezer storage (-20° ± 10°C)

NOTE: PAH – polycyclic aromatic hydrocarbon
PCB – polychlorinated biphenyl
TOC – total organic carbon

[a] Holding times begin the day the sediment sample is collected in the field.
[b] Arsenic, cadmium, chromium, copper, lead , mercury, nickel, selenium, silver, and zinc.
[c] Sample may be held for up to one year if stored at -20°C±10°C (USEPA/USACE 1998).
[d] PCBs as congeners, Aroclors 1242, 1248, 1254, 1260, and total PCBs (USEPA/USACE 1998).
[e] Chlorinated pesticides on USEPA Method 608 list (USACE 1993; USEPA/USACE 1998).
[f] PAH compounds on USEPA Method 610 list (USACE 1993; USEPA/USACE 1998).

Appendix C

Standard Operating Procedures

STANDARD OPERATING PROCEDURE

SEDIMENT CORE/SAMPLE COLLECTION – VIBRACORER

Sediment core samples may be collected with an electrically powered vibracorer, which is lowered through the water column under winch control, and which penetrates the sediment by means of its weight and intense vibration. The following steps outline the procedure for collection of sediment samples using a vibracorer.

1. Maneuver the sampling vessel to the proposed sampling location using the navigation system and deploy a marker buoy at the location.

2. Check to ensure that the metal core barrel is securely fastened to the powerhead of the vibracorer and insert a decontaminated core liner inside the metal core barrel.

3. Insert a core catcher in to the core nose so that the catcher fingers will extend into the core liner, and then screw the core nose onto the bottom of the core barrel.

4. Continue screwing the core nose until the shoulder on the inside of the core nose firmly contacts the end of the core barrel. Tighten the core nose with a spanner or strap wrench.

5. Start the electrical generator, but **DO NOT** energize the corer.

6. Signal the winch operator to hoist the corer and swing it over the stern or side of the vessel at the marked sampling location. Reposition the vessel if necessary. Record the measured water depth, and enter the tidal elevation on the core collection log sheet. Calculate the mudline elevation, and then determine the number of feet of penetration required to reach project depth.

7. Signal the winch operator to lower the corer through the water column. Determine the depth of the corer in the water column and track its subsequent penetration into the sediment either by marking the winch line in 1-ft increments or by attaching a flexible tape measure to the powerhead. In either case, the reference will be 0 ft at the tip of the core nose.

8. When the core nose is within approximately 10 ft of the bottom, energize the corer by actuating the circuit breaker on the generator control panel.

9. Slow the descent speed of the corer in order to determine when the core nose is entering the sediment. Maintain tension on the winch line throughout the coring process to keep the corer from topping over. The worker monitoring the penetration of the corer into the sediment will signal the winch operator when to pay out more line.

10. If refusal is encountered or if the measured distance to the tip of the core nose indicates that project depth has been reached, stop paying out line and de-energize the corer. Do not power down the generator. Refusal is indicated by less than 6 inches of penetration in a given 30-second interval.

11. Signal the winch operator to bring the winch line taut. Maneuver the boom or the boat until the winch pulley is directly above the corer in the sediment, as indicated by the winch line being as close to true vertical as possible.

12. Record the position of the actual coring location. The navigation antenna may be mounted on the winch boom near the pulley to place it directly over the corer.

13. Signal the winch operator to retrieve the corer. If the corer is stuck in the bottom, energize the power head while maintaining tension on the winch line. To reduce the risk of losing sediment from the core barrel, de-energize the corer over the deck and lower it to a holding rack. Note and record the length of smearing on the outside of the core barrel, which gives and indication of the amount of penetration.

14. Use a spanner or strap wrench to unscrew the core nose and remove it. The catcher may stay inside the core nose or remain attached to sediment inside the core liner. Retain any sediment in the core nose and catcher for examination and possible use.

15. Pull the corer liner approximately 6 inches out of the core barrel, remove the catcher (if necessary), and immediately cap the bottom end of the core liner with a plastic cap. Secure the bottom cap with duct tape and proceed to step 16.

    Alternatively, remove the core completely out of the core barrel and evaluate the appearance and length of the core sample by examination through the clear plastic core liner. Note any stratigraphic intervals or other salient features on the core collection log sheet. Extrude the sediment from the core liner and place into food-grade polyethylene bags on board the sampling vessel and proceed to step 25.

16. Extract the core liner entirely from the core barrel, and immediately cap the top of the core liner.

17. If the core is to be cut into length-wise sections, draw a mark on the outside of the core liner where the cut will be made to cut off the bottommost section. Apply duct tape and use a permanent marker to mark the sections on both sides of the location of the future cut. Mark arrows pointing toward the top end of the core, write the core ID, write date and time, and indicate the depth interval spanned by the sections in terms of feet below mudline.

18. Three individuals are needed to complete the cutting process: One person will make the cut with a saw loaded with a decontaminated blade, and two persons will tend the cut ends of the sections.

19. Make the cut and immediately cap both the exposed ends. Immediately secure both caps with duct tape.

20. Repeat the cutting procedure if more length-wise sections need to be cut.

21. Remove the cap from the top end of the top-most section and drain the water. Draining may be accomplished by drilling the hole through the core liner just above the top of the sediment or by gently tipping the section to empty the water out the top. The latter approach may be risky if the sediments are watery or loose.

22. Cut off the excess plastic tube and immediately cap the end and secure the cap with duct tape.

23. If the core will consist of only one section, do steps 15 and 16, mark the core liner as described in step 17, and then do steps 21 and 22.

24. Evaluate the appearance and length of the core sample by examination through the clear plastic core liner. Note any stratigraphic intervals or other salient features on the core collection log sheet.

25. Fill out a chain-of-custody form for the core section(s) to initiate the tracking process.

26. Store the core sections at 4°C (± 2°C) in a refrigerator or iced cooler.

27. Complete any additional entries on the core collection log sheet.

<u>Acceptance criteria for a sediment core sample are as follows:</u>

- The core penetrated to and retained material to project depth or refusal and shows evidence of Merritt Sand.

- Cored material did not extend out the top of the core tube or contact any part of the sampling apparatus at the top of the core tube.

- There are no obstructions in the cored material that might have blocked the subsequent entry of sediment into the core and resulted in incomplete core collection.

If sample acceptance criteria are not achieved, the sample will be rejected. If repeated deployment within 25-50 ft of the proposed location does not result in a sample that meets the appropriate acceptance criteria, the Project Manager will make a decision regarding relocating the proposed sample location.

STANDARD OPERATING PROCEDURE

LABORATORY SEDIMENT CORE/SAMPLE PROCESSING

The following steps outline the general procedure to be followed. The number and subdivisions of berths and composites may vary, depending upon a particular sampling episode.

1. All equipment coming into contact with sediment will be decontaminated before use with each sample to avoid cross contamination.

2. Extrude the sediment from the core liner into a stainless-steel bowl or a 5-gallon high-density polyethylene (HDPE) bucket, depending on the volume.

3. Examine the sediment and record descriptive notes on the core collection log sheet. Parameters may include:
    a. Qualitative sediment description.
    b. Odor
    c. Debris
    d. Biological activity (e.g., detritus, shells tubes, bioturbation, live or dead organisms)
    e. Presence of oil sheen
    f. Any other distinguishing characteristics

4. After the sediment description is complete, homogenize the sediment by hand using a stainless-steel mixing spoon or by using an electric drill with a stainless-steel stirring paddle.

5. Once the sediment has been homogenized, immediately collect a sample for sulfide analysis prior to any other processing. Use a stainless-steel spoon to place sediment into a 4-oz jar. Fill the jar two-thirds full and preserve with one vial of zinc acetate supplied by the analytical laboratory. Immediately screw on the lid, label the jar, and place it in a cooler supplied with ice or frozen blue ice packets.

6. Collect a sample of the homogenized sediment from the individual core for archiving. Fill one 16-oz sample container three-fourths full, screw on the lid, label the jar, and place it in freezer storage for archival purposes.

7. Use aluminum foil or a filtered lid to close the container of homogenized sediment until the remaining cores of the group to be composited for that site have been similarly processed.

8. In a 10-gallon HDPE bucket, combine equal portions of sediment from each individual core of the group to be composited and mix thoroughly (e.g., with an electric drill and stainless-steel paddle) until uniformly homogenized.

9. Collect a sample of the homogenized composite for archiving by filling a 32-oz sample jar three-fourths full, screwing the lid on tightly, labeling the jar, and placing it in freezer storage.

10. Distribute the composited homogenized site sediment to the appropriate sample jars, label the jars, complete the core processing log form and sample tracking form, and place the jars in refrigerated storage for subsequent packing and shipping to analytical laboratories.

11. If it is necessary to archive sediment for possible use in bioassays, ensure that all sample jars for analysis have been filled, then collect two 64-oz glass containers per bioassay.

12. Throughout the sample processing phase, maintain secure storage of sediment and samples; that is, observe proper custody procedures, and continue those procedures until the sample shipping containers are released to the shopping carriers.

13. Any sediment remaining from individual cores that was not used in preparing the homogenized composite should be archived at 4°C for potential subsequent analysis of the individual cores.

Appendix D

Bioassay Standard Test Conditions

Summary of Test Conditions and Test Acceptability Criteria for the Amphipod (*Ampelisca abdita*) 10-Day Sediment Toxicity Test	
1. Test type	Static non-renewal
2. Test duration	10 d
3. Temperature	20 ± 1°C
4. Salinity	20 – 35 ppt
5. Light quality	Ambient Laboratory
6. Light intensity	50 – 100 ft c.
7. Photoperiod	Continuous
8. Test chamber size	1 L
9. Seawater volume	800 mL
10. Sediment depth	40 mm
11. Renewal of seawater	None
12. Age of test organisms	Wild population, immature juveniles
13. # of organisms per test chamber	20
14. # of replicate chambers/concentration	5
15. # of organisms per sediment type	100
16. Feeding regime	None
17. Test chamber cleaning	Lab washing prior to test
18. Test solution aeration	Low bubble (~100/minute)
19. Overlying water	0.45 μm-filtered seawater (at test salinity)
20. Test materials	Test sites, reference and control
21. Dilution series	None
22. Endpoint	% Survival
23. Sample holding requirements	< 8 weeks
24. Sample volume required	4 L
25. Test acceptability criteria	≥ 90% survival in the Control treatment
26. Reference toxicant results	Within 2 SD of laboratory mean

Summary of Test Conditions and Test Acceptability Criteria for the Polychaete (*Neanthes arenacoedentata*) 10-Day Sediment Toxicity Test	
1. Test type	Static-renewal
2. Test duration	10 d
3. Temperature	20 ± 1°C
4. Salinity	20 – 35 ppt
5. Light quality	Ambient Laboratory
6. Light intensity	50 – 100 ft c.
7. Photoperiod	12L/12D
8. Test chamber size	1 L glass beakers
9. Test solution volume	800 L
10. Sediment depth	25 mm (200 mL)
11. Renewal of seawater	None, unless needed. If needed, renew 80% of overlying water at 48 hour intervals
12. Age of test organisms	2-3 weeks
13. # of organisms per test chamber	10
14. # of replicate chambers/concentration	5
15. # of organisms per sediment type	25
16. Feeding regime	None
17. Test chamber cleaning	Lab washing prior to test
18. Test solution aeration	Low bubble (~100/minute)
19. Overlying water	0.45 μm-filtered seawater, at test salinity
20. Test concentrations	Test sites, reference and Control
21. Dilution series	None
22. Endpoint	Survival
23. Sample holding requirements	< 8 weeks
24. Sample volume required	4 L
25. Test acceptability criteria	≥ 90% survival in the Control treatment
26. Reference toxicant results	Within 2 SD of laboratory mean

Summary of Test Conditions and Test Acceptability Criteria for the Mussel (*Mytilus galloprovinciales*) Acute Toxicity Water-Column Test		
1.	Test type	Static non-renewal
2.	Test duration	48 hours
3.	Salinity	28 – 32 ppt
4.	Temperature	16 ± 1°C (mussels)
5.	Light quality	Ambient Laboratory
6.	Light intensity	50 –100 ft c.
7.	Photoperiod	16L/8D
8.	Test chamber size	20 mL vials
9.	Test solution volume	10 mL
10.	Renewal of seawater	None
11.	Age of test organisms	Embryo ≤ 4h old
12.	# of organisms per test chamber	150 – 300
13.	# of replicate chambers per concentration	5
14.	# of organisms per concentration	750 – 1,500
15.	Feeding regime	None
16.	Test chamber cleaning	Lab washing prior to test
17.	Test chamber aeration	None
18.	Elutriate preparation water	Site water
19.	Test concentrations	Test sites, and control
20.	Dilution series	Four concentrations (1, 10, 50, 100%) and a Control.
21.	Dilution water	Natural seawater
22.	Endpoints	%Survival and %normal development
23.	Sampling holding requirements	< 8 weeks
24.	Sample volume required	2L
25.	Test acceptability criteria	≥70% survival and normal development in the controls, <10% abnormal in control

Summary of Test Conditions and Test Acceptability Criteria for the Mysid (*Americamysis bahia*) Acute Toxicity Water-Column Test		
1.	Test type	Static non-renewal
2.	Test duration	96 hours
3.	Salinity	25-30 ppt ± 10 ppt
4.	Temperature	20 ± 1°C
5.	Light quality	Ambient Laboratory
6.	Light intensity	50 –100 ft c.
7.	Photoperiod	16L/8D
8.	Test chamber size	400 mL beaker
9.	Test solution volume	200 mL
10.	Renewal of seawater	None
11.	Age of test organisms	1-5 days; 24 hour range in age
12.	# of organisms per test chamber	10
13.	# of replicate chambers per concentration	5
14.	# of organisms per concentration	50
15.	Feeding regime	daily
16.	Test chamber cleaning	Lab washing prior to test
17.	Test chamber aeration	If needed to maintain >40% saturation
18.	Elutriate preparation water	Site water or Clean sea water
19.	Test concentrations	Test sites, and Lab Control
20.	Dilution series	Four concentrations (1, 10, 50, 100%) and a Lab Control.
21.	Dilution water	Natural seawater/artificial seawater
22.	Endpoints	% Survival
23.	Sampling holding requirements	< 8 weeks
24.	Sample volume required	2L
25.	Test acceptability criteria	≥90% survival in the Lab Controls

Summary of Test Conditions and Test Acceptability Criteria for the Inland Silverside (*Menidia beryllina*) Acute Toxicity Water-Column Test	
1. Test type	Static non-renewal
2. Test duration	96 hours
3. Salinity	5 – 32 ppt ± 10 ppt
4. Temperature	20 ± 1°C
5. Light quality	Ambient Laboratory
6. Light intensity	50 –100 ft c.
7. Photoperiod	16L/8D
8. Test chamber size	400 mL beaker
9. Test solution volume	200 mL
10. Renewal of seawater	None
11. Age of test organisms	9-14 days; 24 hour range in age
12. # of organisms per test chamber	10
13. # of replicate chambers per concentration	5
14. # of organisms per concentration	50
15. Feeding regime	At 48 hrs
16. Test chamber cleaning	Lab washing prior to test
17. Test chamber aeration	If needed to maintain >40% saturation
18. Elutriate preparation water	Site water or Clean sea water
19. Test concentrations	Test sites, and Lab Control
20. Dilution series	Four concentrations (1, 10, 50, 100%) and a Lab Control.
21. Dilution water	Natural seawater/artificial seawater
22. Endpoints	%Survival
23. Sampling holding requirements	< 8 weeks
24. Sample volume required	2L
25. Test acceptability criteria	≥90% survival in the Lab Controls

Summary of Test Conditions and Test Acceptability Criteria for the Bioaccumulation Testing using *Macoma nasuta* and *Nephtys caecoides*	
1. Test type	Static-renewal
2. Test duration	10 or 28-days (compound specific)
3. Salinity	>25 ppt
4. Temperature	12-16 ± 1°C
5. Light quality	Ambient Laboratory
6. Light intensity	50 –100 ft c.
7. Photoperiod	16L/8D
8. Test chamber size	12-L tank
9. Test sediment/test solution volume	4-L sediment/8-L water
10. Renewal of seawater	3x per week
11. Age of test organisms	*Macoma* 2-4 years, 28-45 mm shell length; *Nephtys* large adults
12. # of organisms per test chamber	25 *Macoma*/50 *Nephtys*
13. # of replicate chambers per concentration	5
14. # of organisms per concentration	125 *Macoma*/250 *Nephtys*
15. Feeding regime	None
16. Test chamber cleaning	As needed
17. Test chamber aeration	Moderate as needed
18. Elutriate preparation water	Site water or Clean sea water
19. Test concentrations	Test sediment, reference sediment, and a Lab Control sediment
20. Dilution series	N/A
21. Dilution water	Natural seawater/artificial seawater
22. Endpoints	Bioaccumulation
23. Sampling holding requirements	< 8 weeks
24. Sample volume required	20-L
25. Test acceptability criteria	Adequate mass of organisms at test completion for detection of target analytes